



Genetic diversity and geographical genetic diversity in Colombian accessions of *Lippia alba* (Mill.) N.E. Brown

Diversidad genética y diversidad genética geográfica en accesiones colombianas de *Lippia alba* (Mill.) N.E. Brown

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ARTICLE DATA

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ABSTRACT

Bushy matgrass (*Lippia alba*, Verbenaceae) is a promising plant genetic resource, by their active compounds. The present document studied the molecular genetic diversity and spatial genetic structure of two contrasting populations of *L. alba* in Colombia. Eight RAM, evaluated total DNA of 59 accessions of non-cultivated plants collected in two Colombia regions, Chicamocha and Sumapaz. The expected average heterozygosity (or average heterozygosity genetic diversity of Nei) for the sample had low value ($0.0 \leq H_e = 0.2467 \leq 0.5$). The values of molecular diversity (MD) indicated values in the range of 0.1219 to 0.3425 for seven RAM. The frequency of variants is based on an effective number of alleles [Ae] and expected heterozygosity [H_e], genetic diversity by locus ($h_j = 1 - p^2 - q^2$) had maximum values (near 0.5) in the primers ACA, AG, CGA, and CEC. RAM suitably analyzed *Lippia alba* as an endemic genetic resource. A DNA bank composed of 59 Colombian accessions from *Lippia alba* was set up. The analysis of the spatial global structure shows that the subpopulation Sumapaz is structured, whilst the subpopulation Chicamocha, is in the structuring process. The results suggest in all cases the need for implementing: a) exchange of gene-seed, (b) gene banks with maximum genetic variability and c) induce genetic diversity.

Keywords: anthropochory; GD-GGD; Mantel analysis; Verbena complex; RAM; A-2X DNA protocol extraction.

RESUMEN

El Prontoalivio (*Lippia alba*, Verbenaceae) es un recurso fitogenético promisorio por sus compuestos activos. Este documento determinó la diversidad genética molecular y la estructura genética espacial de dos poblaciones contratantes de *Lippia alba* en Colombia. Ocho RAM, evaluaron el ADN total de 59 accesiones de plantas no cultivadas colectadas en dos regiones colombianas, Chicamocha y Sumapaz. La Heterocigocidad promedio esperada- H_{egg} (Heterocigocidad media o Diversidad genética de Nei) para la muestra de trabajo tuvo un valor bajo ($0,0 \leq H_e = 0,2467 \leq 0,5$). Los valores de diversidad molecular (DM) estuvieron en el rango de 0,1219 a 0,3425 para siete marcadores RAM. Con base en la frecuencia de variantes (Numero efectivo de alelos [Ae] y Heterocigocidad esperada [He], la diversidad genética por locus ($h_j = 1 - p^2 - q^2$) tuvo valores máximos (cerca a

0,5) en los cebadores ACA, AG, CGA y CCA. *Lippia alba* como recurso genético promisorio propio, fue susceptible de ser analizada mediante RAM. Se conformó un banco de ADN con 59 accesiones de *L. alba*. El análisis de la estructura global espacial mostró que la subpoblación Sumapaz está estructurada, mientras que la población Chicamocha está en proceso de estructuración. Los resultados sugieren en todos los casos implementar: a) intercambio de semilla genética, b) bancos de germoplasma de mínimos individuos con máxima variabilidad genética, c) inducir diversidad genética.

Palabras clave: antropocoria; GD-DGG, Mantel analysis; Verbenaceae; RAM; Protocolo de extracción A-2X DNA.

INTRODUCTION

Two groups compose Verbenaceae (to date): a) Verbena complex and b) Verbenaceae (Yuan *et al.*, 2010). Verbena complex is new, formed by three closely related genera, *Verbena* (NA, SA), *Glandularia*, and *Junellia* I and II. *Verbena* and *Glandularia* are monophyletic; the genus *Junellia*, composed of two clades I and II, is non-monophyletic. At the interspecific level Verbena contains two clades NA=North America and SA=South America. *Citharexylum*, *Stachytarpheta*, and the *Lantana/Lippia/Aloysia* complex (Verbenaceae group) are the richest genera in species (Yuan *et al.*, 2010). The Verbenaceae have opposite leaves and flowers with light bilateral symmetry of the corolla; fleshy or dry fruits, usually with two or four seeds divided into two or four segments, include woody trees and shrubs. The genera *Verbena* and *Glandularia* are herbaceous, and the species of *Petrea* are lianas (Yuan *et al.*, 2010).

Lippia alba is used as a natural ingredient in cosmetology (aromatherapy) and the pharmaceutical industry (Pereira *et al.*, 2018). It is one of the most important medicinal plants used, among others, by Brazilian people (Rodrigues *et al.*, 2018). Especially used for its somatic, sedative, antidepressant, and analgesic properties (Pereira *et al.*, 2018; Rodrigues *et al.*, 2018). The essential oil of *L. alba* also has uses as a stomachic, antispasmodic, digestive, anti-hemorrhoid, and anti-asthmatic (de Souza *et al.*, 2019).

The main problem with *Lippia alba* is that its sexual reproduction between and within chemotypes is unlikely, due to its genetic variation, suggesting reproductive isolation (Pierre *et al.*, 2011). In *L. alba*, post-meiotic abnormalities affect the formation of viable pollen grains and are a determining factor of low viability and germination of seeds, presenting important problems in their sexual reproduction (Caetano *et al.*, 2011; Budeguer *et al.*, 2013; Herrera-Moreno *et al.*, 2013).

The asexual reproduction is easy, quick and successful. Additionally, in Colombia the species is subject to a high extractive regime without replacement, causing genetic erosion, being an unattended species without conservation plans-programs.

The genetic structure of natural populations of the genus *Lippia* has been barely studied (Reis *et al.*, 2014). Previous analyzes using RAPD markers were effective to understand the genetic diversity of species of *Lippia* spp., and contributed to understanding his adaptation to the environment, conservation, and taxonomic implications (Gomide *et al.*, 2013). Martínez-Natarén *et al.*, (2014), evaluated the degree of genetic diversity in *Lippia graveolens* in wild populations of Mexican oregano; Viccini *et al.* (2014) assessed the possible association between the variations in the production of secondary metabolites and genetic traits in 37 accesiones of *Lippia alba* from various regions in Brazil. dos Santos *et al.* (2015) to evaluate the chemical diversity of *L. sidoides* genotypes

and determined that the chemical polymorphism concerning the compounds thymol and carvacrol was caused by genotypes.

Microsatellite primers were developed in Brazil and optimized for *Lippia alba* to characterize the *L. alba* germplasm by Rocha *et al.* (2015). The data provide support to characterize germplasm banks, genetic breeding programs for *L. alba*, and other genetic diversity studies and classifications of species in the genus *Lippia*. In Brazil, do Amaral *et al.* (2017) obtained 74/97 polymorphic bands in *L. alba*, confirming genetics and phytochemistry variability among genotypes of the same region using ISSR markers. In *L. sodoides* and *L. gracilis*, 11/20 microsatellites markers showed polymorphism, proving its effectiveness in assessing the genetic diversity of promissory species (Santos *et al.*, 2014).

Random Amplified Markers-RAM (Ng and Tan, 2015; Grover and Sharma, 2016), has been evaluated in *Prunus dulcis* (Rasouli *et al.*, 2015), *Opuntia focus indica* (Zarroug *et al.*, 2015), *Helianthus annuus* (Sala *et al.*, 2017), fungal biodiversity (Shamim *et al.*, 2017), the population structure of *Hymenoscyphus fraxineus* (Burokiene *et al.*, 2015), genetic variation in *Phlebiopsis gigantea* (Dar *et al.*, 2017) and phytopathogens (Gharbi *et al.*, 2014; Nath *et al.*, 2016) in species of economic importance.

The research was designed to obtain estimates of the genetic diversity and spatial genetic structure of *Lippia alba* in Colombia, to determine: a) the relationships between genetic and spatial distances and b) the factors that cause changes in the amount of variation between populations and

localities; in two contrasting populations of *L. alba* in Colombia: Chicamocha (Tropical dry forest) and Sumapaz (Tropical humid forest).

MATERIALS AND METHODS

Fifty-nine introductions of *Lippia alba* were collected from different localities in two Colombian ecological zones; Region I: Sumapaz [4,3124166N, -74,493694W], with 35 collections; and Region II: Chicamocha [6,5683333N, -73,1400277W], with 24 collections. Four individuals per introduction made up the work sample. These introductions were part of a germplasm bank of a transitory collection *in vivo* and *ex situ* at the National University of Colombia-Palmira. Details of collection data in Cardona (2014).

DNA extraction. The protocol 'A-2X' of DNA extraction for aromatic species developed by Vega-Vela and Chacon (2011) was used. Total DNA was extracted from 100-200 mg of fresh leaf tissue from each introduction by the A-2X method. The quality and concentration of the DNA were evaluated in a 0.8% agarose gel. DNA integrity was confirmed by 0.8% TBE buffer polyacrylamide gel electrophoresis.

RAM technics. PCR was performed for the primers ACA, AG, CA, CCA, TG, CT, CGA, and GT. PCR products were visualized in 7% polyacrylamide gels (37:1) at 160 volts for one hour and Ethidium Bromide staining was done. The mixture for the PCR was: DNA 10.0 ng/ μ L + TAQ buffer 10X 2.50 μ L + MgCl₂ 2.50. Table 1 shows the amplification protocol used.

Table 1. Amplification protocol with temperature and time by Step-Stage Standardized PCR.

Step	T°	t (time)	Stage
1	95	5 minutes	Denaturalization
2	95	30 segundos	Desnaturalization
3	HT*	45 segundos	Hybridization
4	72	2 minutes	Length
5	37	Times from step 2	
6	72	7 minutes	Length
7	16	30 minutes	

Primer	HT*
ACA-AG-CA	50
CCA-TG-CT	55
GT-CCA	58

Data analysis. The bands were characterized by the presence/absence in each introduction and the data were entered in a binary matrix 1/0. The allelic and genotypic frequencies (observed and expected); and the values of genetic diversity by locus ($h_j=1-p^2-q^2$) and average ($H_i=\sum h_j/n$) of Nei (1987), were calculated using the Program TFPGA v1.3. Potential discrimination of each primer or degree of genetic variability in the population was expressed by the Simpson Coefficient ($H=\sum (1-\sum p_i^2)/n$), where p_i is the frequency of the i^{th} allele and n is the number of loci detected by each primer (Manica-Cattani *et al.*, 2009). For all pairs of genotypes, the genetic similarity values were calculated using the Dice-Nei coefficient (Kosman and Leonard, 2005), which excludes the 0-0 value as an indicator of similarity (Manica-Cattani *et al.*, 2009). To determine the level of genetic differentiation, the GenAlEx program was used (Peakall & Smouse, 2006). In addition, four statisticians were estimated based on the specific number of alleles per locus within accession.

RESULTS AND DISCUSSION

Nei's genetic diversity value for the sample was low ($D=0.2467$). Molecular diversity estimates for seven RAM ranged from 0.1219 - 0.3485 (Table 2). Seven of eight primers amplified, generating consistent and reproducible bands at 94 loci. The expected Heterogeneity was 0.4008. Each primer generated 13.43 ± 4.43 average loci (Table 2), with $P_i=91.49\%$ of polymorphic loci. The ACA, AG, CGA, and CCA primers showed high values of genetic diversity per locus (h_j), close to the maximum of 0.5. The Discriminant-H-values of the primers ranged from 0.1755 to 0.9813. The allele frequencies took all possible values between 0 and 1, with important variations between and within primers. The averages for the statisticians were specific: $n_x=25.97$; $Ae_x=2.4062$; $P_x=0.9260$; $Ho_x=0.4008$; $He_x=0.3595$; and the $DG_x=0.2467$ (Table 2). The number of alleles individually in each amplified sequence can be displayed in Cardona (2014).

Table 2. The number of loci, polymorphisms, and values of genetic diversity.

Primer	Seq.	LOCI		DG (Nei, 1987)							INTRODUCTIONS				
		No. Loci	Poly	AMONG			WIHTING				Multi	Mono			
				DG	N	Ae	P	Ho	He	Hi	Pi	Hi			
1	ACA	19	19	0	0.3339	26.42	3.7245	1.0000	0.4478	0.7315	0.3603	0.9831	0.7656	58	1
2	AG	14	14	0	0.3425	37.07	0.1427	1.0000	0.6283	0.4994	0.4298	0.9661	0.5634	57	2
3	CA	19	13	6	0.1219	49.32	0.0638	0.6842	0.8359	0.1755	0.4776	1.0000	0.2991	59	0
4	CGA	9	8	1	0.1718	11.89	1.3930	0.8889	0.2015	0.9202	0.1765	0.7966	0.9270	47	12
5	CCA	8	8	0	0.3317	23.63	6.6881	1.0000	0.1250	0.9813	0.3252	0.9322	0.8001	55	4
6	TG	11	10	1	0.2483	22.45	0.4339	0.9091	0.3806	-1.3048	0.1363	1.0000	0.9632	59	0
7	GT	14	14	0	0.1768	11.00	2.0549	1.0000	0.1864	0.5134	0.0000	0.0000	0.8136	0	59

Specific Statisticians (N=average number of alleles/locus, Ae=effective number of alleles, P=proportion of polymorphic loci, Ho= heterozygosity observed, He=expected heterozygosity), and statisticians by accession (Pi=Proportion of accessions multilocus, Hi= average heterozygosity by accession).

Eighty-six of 94 loci were found to be polymorphic. The number of loci by primer was in your order ACA>CA>AG>GT>TG>CGA>CEC and the proportion of polymorphic loci was high (0,907).

Molecular genetic diversity estimated had an intermediate value on the 0-0,5 Nei's scale for RAM. Values distributed across the minimum-maximum range $0.167 \leq Mgd \leq 0.332$ is rated as 'medium'. These values are common in Colombia for *L. alba* and *Lippia origanoides* H.B.K (Suárez *et al.*, 2007; Suárez *et al.*, 2008). The values of genetic variability for *L. origanoides* ($H=0.453$) in Colombia estimated by Suárez *et al.*, (2008), were lower than those found by Manica-Cattani *et al.* (2009) among *L. alba* accessions collected in southern Brazil, with mean values of 0.565 for ISSR and 0.625 for RAPD.

The degree of genetic variability was high ($H=0.7356$). The potential discrimination of each primer expressed by the Simpson Coefficient showed values between 0.2991 for the AG primer, to 0.9632 for the TG primer. However, for species such as *L. alba* and *L. origanoides*, these values ($H=0.484$) are common in Colombia (Suárez *et al.*, 2007; Suárez *et al.*, 2008). On the other hand, Manica-Cattani *et al.* (2009) found high genetic variability among *L. alba* introductions collected in southern Brazil, with average values of 0.565 for ISSR and 0.625 for RAPD.

In the sample job, the allelic frequencies showed all possible values within the range, a factor that can be considered characteristic of RAM. Different from the frequencies obtained by Manica-Cattani *et al.* (2009), with extreme high (>0.90) and low (<0.10) values in Brazilian accessions with the same type of dominant marker (RAPD and ISSR). The discriminating power of RAMs has been valued in local plant genetic resources including heliconias (Arcos *et al.*, 2004), native trees of *Psidium guajava* (Muñoz *et al.*, 2008; Sanabria *et al.*, 2006), *Rubus* spp (Muñoz *et al.*, 2008), cape gooseberry (Bonilla *et al.*, 2008).

Viccini *et al.* (2014), evaluated the degree of genetic diversity in nine *Lippia* species such as (*L. corymbosa*, *L. diamantinensis*, *L. filifolia*, *L. florida*, *L. hermannioides*, *L. lupulina*, *L. rotundifolia*, *L. rosella* and *L. sidoides*) from southwestern Brazil. The average interspecific genetic distance was similar for all species and higher than the intraspecific distances. Species with limited presence showed lower interspecific diversity. The dendrogram by the UPGMA method showed larger groups with a clear differentiation between species (Viccini *et al.*, 2014).

Suárez *et al.* (2007) conducted a pilot study in *Lippia origanoides*, *L. alba*, and *L. citriodora* to observe intra and inter specific polymorphisms in four regions of cpDNA, ribosomal DNA STIs, and 50 ISSR loci. Three of four cpDNA regions (petA-psbE, trnL-trnF and trnL intron) and the ITS region showed mostly interspecific variation. Seventeen ISSR regions from 50 loci analyzed showed polymorphisms, both inter and intraspecific. Sequence divergence between species pairs in the plastid region showed ranges of 0.4% to 1%, while the divergence in ITS region sequences was about 5%. The results obtained suggest that the STI regions, as reported for other plant genera, may become a region of choice for population genetics studies in this type of species. The genetic diversity values described by Suárez *et al.* (2008) in *Lippia origanoides* (related Verbenaceae cross-pollinated plants) were for percentage of polymorphic loci 86-21%; Shannon diversity index of 0-453; and for average panmitic heterocigosity of 0-484.

Genetics differentiation level between populations. Analysis of molecular variance-AMOVA and F_{test} , showed the lowest haplotypic variation among the population (*Est.Var. AP=0,141*), with remaining (95%) distributed within the population (*Est.Var.WP=2,46*). The value F_{PT} was 0.054 ($p<0.001$), signaling small genetic differentiation between regions (Table 3).

Table 3. AMOVA and F_{PT} test values for 58 Colombian accessions *L. alba*.

Source	Df	SS	MS	Est. Var.	Proportion
Among Pops	1	6.381	6.381	0.141	5%
Within Pops	56	137.744	2.460	2.460	95%
Total	57	144.125		2.601	100%

Stat	Value	P(rand \geq data)
PhiPT	0.054	0.001
PhiPT max	0.948	
Phi'PT	0.057	

Principal Coordinates Analysis-ACP, AMOVA, and Bayesian analysis conducted by Suárez *et al.* (2008) on Colombian populations of *L. origanoides* revealed a low level of genetic differentiation between two localities within the Chicamocha Canyon-Colombia, suggesting a single population of *Lippia origanoides*. The levels of Genetic Diversity-GD in this population expressed as a percentage of polymorphic loci ($P=86.21\%$), the Shannon diversity index ($I=0.453$) and the average panmitic heterocigocity ($HB=0.484$), are comparable to the GD levels of other related cross-pollinated Verbenaceae (Suárez *et al.*, 2008).

The similarity matrix between the two groups, based on the Nei and Li coefficient, PCoA and Mantel analysis (Figures 1 and Figure 2), revealed the existence of different geographic clusters in each subpopulation. The level of differentiation between the two populations was relatively low but significant. Martínez *et al.* (2008) obtained the same result in *L. origanoides*, a related species, in the Chicamocha-Colombia region. Mantel analysis grouped individuals by their characteristics or geographical proximity. Spatial autocorrelation analyses showed a consistent pattern of isolation by distance with a moderate but significant level of spatial structure.

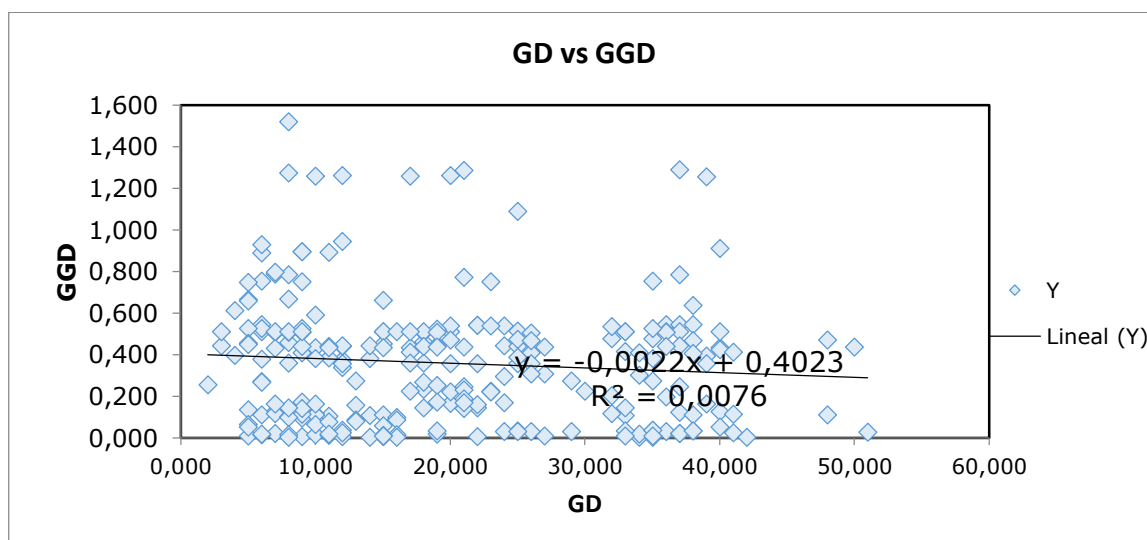


Figure 1. Region_1: Chicamocha. Mantel results for Genetic Diversity-GD vs Geographical Genetic Diversity-GGD.

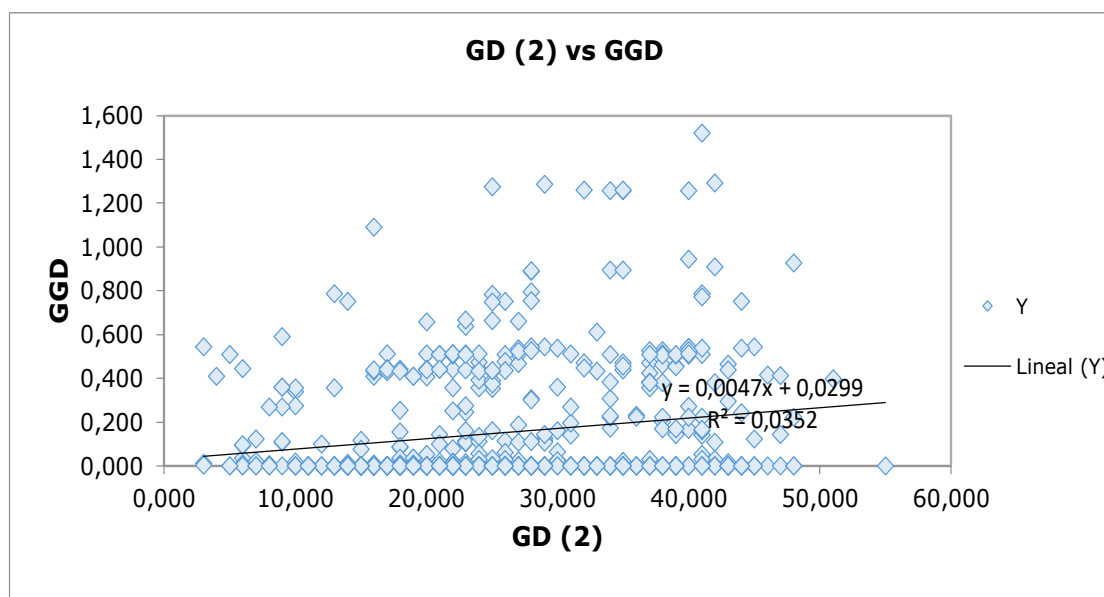


Figure 2. Region_2: Sumapaz. Mantel results for Genetic Diversity-GD vs Geographical Genetic Diversity-GGD.

These results suggest serve as proposed by Suárez *et al.* (2008), those who refer to sample this type of Verbenaceae at distances greater than 1.2 km obtaining different genotypes, which would help preserve GD levels. The causes of this spatial pattern are unknown and may be associated, among others, to restricted dispersal of seed or pollen movement short distances (Suárez *et al.*, 2008).

Spatial structure. The analysis of the spatial global structure shows that the subpopulation Sumapaz is structured, whilst the subpopulation Chicamocha, is in structuring process. The relationship between geographic distances and distances of Nei was significant ($p \leq 0.001$), indicating isolation by distance between the subpopulations (Figure

3 and Figure 4). The relationship between geographic distance and genetic distance, was significant for Sumapaz region ($p \leq 0.04$) and non-significant for Chicamocha region ($p \geq 0.23$); indicating isolation by distance in Sumapaz and not isolation by distance in Chicamocha. Figure 3 show the spatial structure between populations. The correlogram ($\Omega = 97.136$) for the subpopulation Sumapaz showed spatial structure significant [$0.001 P(\Omega\text{-rand} \geq \Omega\text{-data})$]. The correlogram ($\Omega = 28.488$) to the spatial structure of the population Chicamocha was not significant [$0.098 P(\Omega\text{-rand} \geq \Omega\text{-data})$]. The overall spatial structure analysis shows that the subpopulation Sumapaz is structured, while that Chicamocha, no.

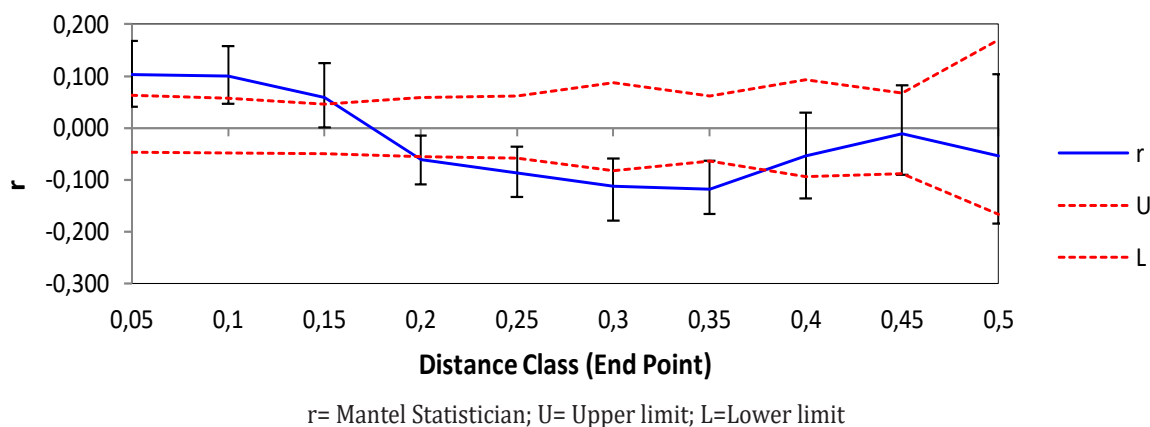


Figure 3. Pooled spatial structure for the two populations (Chicamocha and Sumapaz).

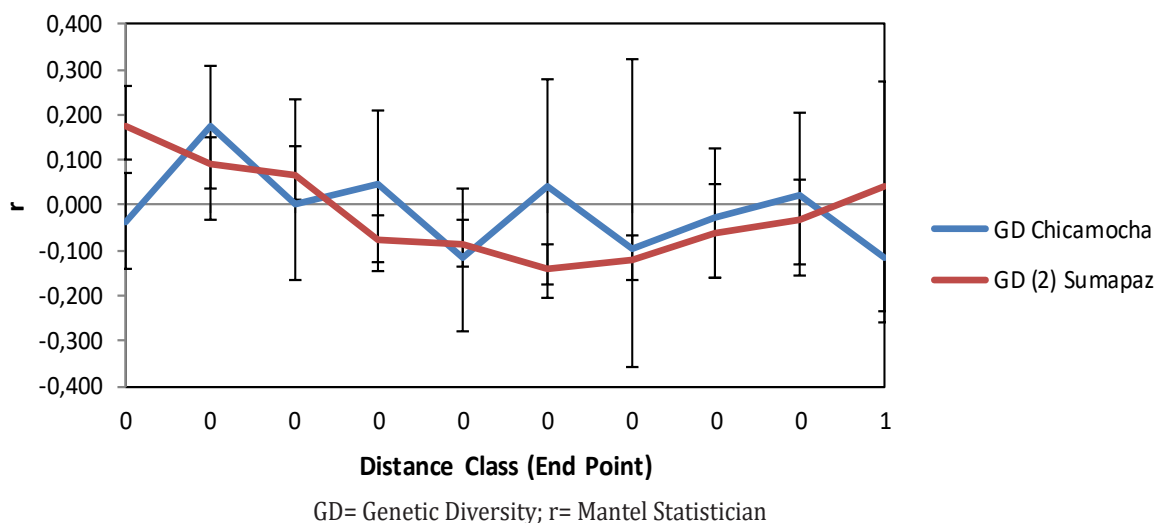


Figure 4. Spatial structure for each sub-population, Chicamocha y Sumapaz, based on Genetic Diversity-GD.

Biologically, population structuring processes are attributable to: (i) contrasting local environmental conditions and (ii) use-consumption as anthropic factor, (iii) genetic factors, (iv) possible polyploidies, (v) chemotypes (citral, carvone, linalool), (vi) reproductive isolation, (vii) inviable pollen. Contrasting environmental conditions such as Bs-T in Chicamocha and Bh-T in Sumapaz; and increased use-consumption of the species in the Chicamocha sub-region, are factors causing changes in the amount of variation between

the two populations in each locality; and were decisive in the geographical distribution of the genetic diversity found. In the Chicamocha sub-region a greater number of repeated biotypes and chemotypes were found (Cardona, 2014), causing reproductive isolation by mixoploidia. Lower biomass and distance between collections were also observed. However, the biotypes collected in the Chicamocha sub-region showed higher content and quality of essential oils (Cardona, 2014).

Four factors that may also be affecting genetic diversity of the populations studied: a) post meiotic abnormalities, determining factor of both low viability and germination of seeds (Reis *et al.*, 2014), having breathing problems of mating (Venâncio *et al.*, 2016); b) anthropochory (other way of dissemination), c) stenopropagation (vegetative propagation quick and easy) and, d) extractive regime without replacement. Further, restricted seed spread and short distances of pollen movement, proposed by Venâncio *et al.* (2016).

CONCLUSIONS

In the two sub-populations studied, low genetic diversity and structure were detected. The low genetic diversity found could be based on: i) a significant number of Carvona chemotypes included in the working sample, ii) possible polyploidies (not studied), iii) the creation of reproductive isolation between populations, and of different individuals in the population.

The methodology used allowed us to obtain values of genetic diversity and genetic population structure for each of the populations studied; these values are within the range for different species of the genus *Lippia* and agree with the common values reported in Colombia by other authors.

Intermediate levels of DG indicate recent isolations, which prevent the development of strong genetic structure of each subpopulation; or low or no gene flow between individuals in each region. Possible barriers are biological with a negative impact on the genetic composition of *L. alba*. The loss of DG has had an enormous negative impact on the evolutionary potential of the species.

The result obtained here and by other authors, allows us to affirm that the Colombian territory

is part of the center of the genetic diversity or primary natural distribution of the species.

Lippia alba as a promising plant genetic resource was susceptible to analysis by the RAM. The discriminant power of the ARS was effective in *L. alba* based on the results generated in other studies on the species (*L. alba*) and related species.

BIBLIOGRAPHIC REFERENCES

- Arcos, A. L.; Mondragón, A. J.; Muñoz, J. E.; Botero, S. (2004). Colecta y caracterización molecular con marcadores tipo RAM (Microsatélites aleatorios) de heliconias y especies relacionadas. Colombia: Congreso Colombiano de Botánica: Botánica, Diversidad y Cultura.
- Bonilla, M. L.; Piedrahita, K. E.; Terranova, A. M. P.; Amariles, H. D. V.; Flórez, J. E. M. (2008). Caracterización molecular de 43 accesiones de uchuva de seis departamentos de Colombia. *Acta Agronómica*. 57(2): 109-115.
- Budeguer, C. J.; Nasif, A.; Martínez Pulido, L.; Pastoriza, A.; Andrada Mansilla, B. (2013). Cytogenetics of *Lippia alba* (Mill.) Brown from Lules, Tucumán. *Revista Agronómica del Noroeste Argentino*. 33(1): 11-14.
- Burokiene, D.; Prospero, S.; Jung, E.; Marciulyniene, D.; Moosbrugger, K.; Norkute, G.; Rigling, D.; Lygis, V.; Schoebel, C. N. (2015). Genetic population structure of the invasive ash dieback pathogen *Hymenoscyphus fraxineus* in its expanding range. *Biological Invasions*. 17(9): 2743-2756.
- Caetano, C. M.; Serna, C.; Yamil, D.; Muñoz Galindez, E. (2011). 10 años de la maestría en ciencias biológicas, línea de investigación en recursos Fitogenéticos Neotropicales: formando capacidades para contribuir a la conservación del patrimonio genético y la valoración del conocimiento tradicional. Recovered from <https://repositorio.unal.edu.co/handle/unal/8361>
- Cardona, J. O. (2014). *Estructura genética y fitogeografía de poblaciones colombianas de Lippia alba (Mill.) NE Brown (Verbenaceae)*. Recovered from <http://www.bdigital.unal.edu.co/12886/>

- Dar, R. A.; Saba, I.; Shah Nawaz, M.; Qazi, P. H.; Khan, I. A. (2017). Antimicrobial potential of fungal endophytes from selected high value medicinal plants of the Kashmir valley-India. *The Journal of Psychopharmacology*. 6(5): 307-310.
- de Souza, E. M.; de Souza, R. C.; Melo, J. F.; da Costa, M. M.; de Souza, A. M.; Copatti, C. E. (2019). Evaluation of the effects of *Ocimum basilicum* essential oil in Nile tilapia diet: growth, biochemical, intestinal enzymes, hematology, lysozyme and antimicrobial challenges. *Aquaculture*. 504: 7-12.
- do Amaral, U.; Pereira, M. B.; Junior, P.; de Souza, M.; Bajay, M. (2017). Polymorphism in *Lippia alba* clones from the metropolitan region of Rio de Janeiro. *Journal of Advance in Agriculture*. 7(2): 1044- 1049.
- dos Santos, C. P.; de Oliveira, T. C.; Pinto, J.; Fontes, S.; Cruz, E.; Arrigoni-Blank, M.; Andrade, T.; de Matos, L. Alves, P.; Innecco, R.; Blank, A. F. (2015). Chemical diversity and influence of plant age on the essential oil from *Lippia sidoides* Cham. germplasm. *Industrial Crops and Products*. 76: 416-421.
- Gharbi, Y.; Triki, M. A.; Jolodar, A.; Trabelsi, R.; Gdoura, R.; Daayf, F. (2014). Genetic diversity of *Verticillium dahliae* from olive trees in Tunisia based on RAMS and IGS-RFLP analyses. *Canadian journal of plant pathology*. 36(4): 491-500.
- Gomide, M. D.; Lemos, F. D. O.; Lopes, M. T.; Tânia, M. D.; Viccini, L.; Coelho, C. M. (2013). The effect of the essential oils from five different *Lippia* species on the viability of tumor cell lines. *Revista Brasileira de Farmacognosia*. 23(6): 895-902.
- Grover, A.; Sharma, P. C. (2016). Development and use of molecular markers: past and present. *Critical reviews in biotechnology*. 36(2): 290-302.
- Herrera-Moreno, A. M.; Carranza, C. E.; Chacón-Sánchez, M. I. (2013). Establishment of propagation methods for growing promising aromatic plant species of the *Lippia* (Verbenaceae) and *Tagetes* (Asteraceae) genera in Colombia. *Agronomía Colombiana*. 31(1): 27-37.
- Kosman, E.; Leonard, K. J. (2005). Similarity coefficients for molecular markers in studies of genetic relationships between individuals for haploid, diploid, and polyploidy species. *Molecular Ecology*. 14(2): 415-424.
- Manica-Cattani, M. F.; Zacaria, J.; Pauletti, G.; Atti-Serafini, L.; Echeverrigaray, S. (2009). Genetic variation among South Brazilian accessions of *Lippia alba* Mill. (Verbenaceae) detected by ISSR and RAPD markers. *Braz. J. Biol.* 69(3): 375-380.
- Martínez-Natarén, D. A.; Parra-Tabla, V.; Ferrer-Ortega, M. M.; Calvo-Irabién, L. M. (2014). Genetic diversity and genetic structure in wild populations of Mexican oregano (*Lippia graveolens* HBK) and its relationship with the chemical composition of the essential oil. *Plant systematics and evolution*. 300(3): 535-547.
- Martinez, F. (2008). Genetic Diversity Within and Among Wild and Garden Aromatic Species of the Genera *Lippia*, *Aloysia* and *Phyla* in Several Locations in Northeastern Colombia. Recovered from <http://noesis.uis.edu.co/bitstream/123456789/30345/1/140155.pdf>
- Muñoz, J. E.; Coronado, A. C.; Coronado, Y. M. (2008). Microsatélites amplificados al azar (RAM) en estudios de diversidad genética vegetal. *Acta agronómica*. 57(4): 219-226.
- Nath, V. S.; Basheer, S.; Jeeva, M. L.; Veena, S. S. (2016). Genetic and Phenotypic characterization of *Phytophthora colocasiae* in Taro Growing Areas of India. *J. Plant Pathol Microbiol*. 7(383): 2-7.
- Ng, W.; Tan, S. G. (2015). Inter-simple sequence repeat (ISSR) markers: are we doing it right. *ASM Sci J*. 9(1): 30-39.
- Peakall, R. O.; Smouse, P. E. (2006). GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. *Molecular Ecology Resources*. 6(1): 288-295.
- Pereira, I.; Aleksandrazielińska, F. J. V.; Santos, A.; Nowak, I.; Silva, A.; Souto, E. B. (2018). Monoterpenes-Based Pharmaceuticals: A Review of Applications in Human Health and Drug Delivery Systems. In: Goyal, M.; Chauhan, D. *Plant-and Marine-Based Phytochemicals for Human Health: Attributes, Potential, and Use*. pp. 85-130. Canada: CRC Press. 368p.

- Pierre, P. M.; Sousa, S. M.; Davide, L. C.; Machado, M. A.; Viccini, L. F. (2011). Karyotype analysis, DNA content and molecular screening in *Lippia alba* (Verbenaceae). *Anais da Academia Brasileira de Ciências*. 83(3): 993-1006.
- Rasouli, M.; Martínez, G. P.; Karimi, R. (2015). Application of Random Amplified Microsatellite Polymorphism (RAMP) in *Prunus* characterization and mapping. *Journal of Nuts*. 6(01): 1-5.
- Reis, A. C.; Sousa, S. M.; Vale, A. A.; Pierre, P. M.; Franco, A. L.; Campos, J. M.; Vieira, R. F.; Viccini, L. F. (2014). *Lippia alba* (Verbenaceae): A new tropical autopolyploid complex? *American Journal of Botany*. 101(6): 1002-1012. doi:10.3732/ajb.1400149
- Rocha, D. S.; Santos, C. P. D.; Bajay, M. M.; Campos, J. B. D.; Blank, A. F.; Pinheiro, J. B.; Zucchi, M. I. (2015). Development of a novel set of microsatellite markers for *Lippia alba* (Verbenaceae). Recovered from <https://www.repositorio.ufs.br/handle/riufs/7289>
- Rodrigues, R. A. F.; Figueira, G. M.; Sartoratto, A.; Yamane, L. T.; de Freitas-Blanco, V. S. (2018). Chemical Diversity and Ethno pharmacological Survey of South American Medicinal and Aromatic Plant Species. In: *Medicinal and Aromatic Plants of South America*. pp. 17-44. Dordrecht: Springer.
- Sala, C.; Ramos, M. L.; Bulos, M.; Altieri, E. (2017). *U.S. Patent No. 9,574,237*. Washington, DC: U.S. Patent and Trademark Office.
- Shamim, M.; Kumar, P.; Kumar, R. R.; Kumar, M.; Kumar, R. R.; Singh, K. N. (2017). Assessing Fungal Biodiversity Using Molecular Markers. In: *Molecular Markers in Mycology*. pp. 305-333. Cham: Springer,
- Sanabria, H. L. (2006). Caracterización molecular con marcadores RAM de árboles nativos de *Psidium guajava* (guayaba) en el Valle del Cauca. *Acta Agronómica*. 55(1): 23-30.
- Santos, C. P.; Rocha, D. S.; Bajay, M. M.; Santos, F. R. C.; Campos, J. B.; Pinheiro, J. B.; Zucchi, M.; Silva-Mann, M.; Arrigoni-Blank, M.; Blank, A. F. (2014). Cross-species transferability of microsatellite markers in the genus *Lippia*. *Genetics and Molecular Research*. 13(4): 9846-9850.
- Suárez, A R.; Martínez, F. O.; Núñez, G. A.; Castillo-Villamizar, G. A.; Chacón M I. (2007). Molecular characterization of aromatic species of the genus *Lippia* from the Colombian neotropics, Recovered from https://www.actahort.org/books/756/756_14.htm
- Suárez, A R.; Castillo, G.A.; Chacón, M. I. (2008). Genetic diversity and spatial genetic structure within a population of an aromatic shrub, *Lippia origanoides* (Verbenaceae), in the Chicamocha Canyon, northeastern Colombia, *Genetics Research*. 90(6): 455-465.
- Venâncio, D.; Viccini, L.; Luizi-Ponzo, A.; Prezoto, F. (2016). Flower-Visiting Insects and Phenology of *Lippia alba* (Lamiales: Verbenaceae): Floral Color Changes and Environmental Conditions as Cues for Pollinators. *Environmental entomology*. 45(3): 685-693.
- Vega-Vela, N. E.; Chacón, M. I. (2011). Isolation of high-quality DNA in 16 aromatic and medicinal Colombian species using silica-based extraction columns. *Agronomía Colombiana*. 29(3): 349-357.
- Viccini, L.F., Silveira, R.S.; do Valea, A.A.; de Campos, J.M.; Reis, A.C.; de Oliveira Santos, M.; Campos, V.R.; Carpane, A.G.; Grazul, R.M. (2014). Citral and linalool content has been correlated to DNA content in *Lippia alba* (Mill.) NE Brown (Verbenaceae). *Industrial Crops and Products*. 59: 14-19.
- Yuan, Y. W.; Liu, C.; Marx, H. E.; Olmstead, R. G. (2010). An empirical demonstration of using pentatricopeptide repeat (PPR) genes as plant phylogenetic tools: Phylogeny of Verbenaceae and the Verbena complex. *Molecular Phylogenetics and Evolution*. 54(1): 23-35.
- Zarroug, M. B.; Baraket, G.; Zourgui, L.; Souid, S.; Hannachi, A. S. (2015). Genetic diversity and phylogenetic relationship among Tunisian cactus species (*Opuntia*) as revealed by random amplified microsatellite polymorphism markers. *Genetics and Molecular Research*. 14(1): 1423-1433.