



# *Trichoderma asperellum* (Samuels, Lieckf & Nirenberg) as a promoter of vegetative growth in soybeans

*Trichoderma asperellum* (Samuels, Lieckf & Nirenberg) como promotor del crecimiento vegetativo en soja

Aloisio Freitas Chagas Junior<sup>1</sup>; Lillian França Borges Chagas<sup>2</sup>; Brigitte Sthepani Orozco Colonia<sup>3</sup>; Albert Lennon Lima Martins<sup>4</sup>

## AUTHORS DATA

1. Dr. Agronomy Course, Federal University of Tocantins/UFT/PPGPV, Campus de Gurupi, Gurupi, TO, Brazil, [chagasjraf@mail.uft.edu.br](mailto:chagasjraf@mail.uft.edu.br), <https://orcid.org/0000-0002-7489-8701>
2. Dr. Agronomy Course, Federal University of Tocantins/UFT, Campus de Gurupi, Gurupi, TO, Brazil, [lillianfbc@mail.uft.edu.br](mailto:lillianfbc@mail.uft.edu.br), <https://orcid.org/0000-0002-0083-6452>
3. PhD student, Federal University of Paraná, Curitiba, PR, Brazil, [bricolonia@gmail.com](mailto:bricolonia@gmail.com), <https://orcid.org/0000-0001-8228-3101>
4. Dr. Agronomy Course, State university of Tocantins/Unitins, Palmas, TO, Brazil, [eng.albertlennon@gmail.com](mailto:eng.albertlennon@gmail.com), <https://orcid.org/0000-0003-2683-2035>



Cite: Chagas Junior, A.F.; Chagas, L.F.B.; Colonia, B.S.O.; Martins, A.L.L. (2023). *Trichoderma asperellum* (Samuels, Lieckf & Nirenberg) as a promoter of vegetative growth in soybeans. *Revista de Ciencias Agrícolas*. 39(E): 50-68. <https://doi.org/10.22267/rcia.202239E.199>

Received: June 09 2021. Accepted: December 18 2022.

## ABSTRACT

With the intensification of the problems encountered in agriculture, the use of microorganisms shows promise. Fungi of the *Trichoderma* genus are found naturally in the soil and have been; they are used as an active ingredient in biofungicides and have activity as promoters of plant growth. This work aimed to evaluate the efficiency of the commercial product TrichoPlus, containing *Trichoderma asperellum* (Samuels, Lieckf & Nirenberg) as a promoter of soybean plant growth. Four independent experiments were carried out in different periods between 2019 and 2020. The experiments were conducted in a greenhouse, and biomass and nutrient contents were examined. In the first experiment, among the different rates of TrichoPlus tested (3, 4, 5, and 6g per kg of seeds), the best dose for most of the characteristics evaluated was 5g per kg of seed. Hence, the inoculation of TrichoPlus at 5g per kg of seed in the other experiments showed positive results in the accumulation of biomass, nitrogen, and phosphorus content. Based on the study's results, the product TrichoPlus, composed of *T. asperellum*, can be used as an efficient inoculant for promoting soybean plant growth.

**Keywords:** Bioinoculant; fungus; *Trichoderma*; *Glycine max* (L.) Merrill; biomass; nutrients.

## RESUMEN

Con la intensificación de los problemas encontrados en la agricultura, el uso de microorganismos se muestra prometedor. Los hongos del género *Trichoderma* se encuentran naturalmente en el suelo; se utilizan como ingrediente activo en biofungicidas y tienen actividad como promotores del crecimiento vegetal. Este trabajo tuvo como objetivo evaluar la eficiencia del producto comercial TrichoPlus, que contiene *Trichoderma asperellum* (Samuels, Lieckf & Nirenberg) como promotor del crecimiento de

plantas de soja. Se llevaron a cabo cuatro experimentos independientes en diferentes períodos entre 2019 y 2020. Los experimentos se realizaron en un invernadero y se examinaron los contenidos de biomasa y nutrientes. En el primer experimento, entre las diferentes dosis de TrichoPlus probadas (3, 4, 5 y 6g por kg de semilla), la mejor dosis para la mayoría de las características evaluadas fue de 5g por kg de semilla. Por lo tanto, la inoculación de TrichoPlus a 5g por kg de semilla en los otros experimentos mostró resultados positivos en la acumulación de contenido de biomasa, nitrógeno y fósforo. Con base en los resultados del estudio, el producto TrichoPlus, compuesto por *T. asperellum*, puede ser utilizado como un inoculante eficiente para promover el crecimiento de las plantas de soja.

**Palabras clave:** Bioinoculante; hongo; *Trichoderma*, *Glycine max* (L.) Merrill; biomasa; nutrientes.

## INTRODUCTION

The high productivity of crops such as soybean (*Glycine max* (L.) Merrill) is generally associated with high doses of fertilizers, among other factors. These, in turn, represent a significant portion of the production costs of the crop, in addition to being obtained from a non-renewable source and potential environmental pollutants. The use of plant growth-promoting microorganisms in significant agricultural commodities has been gaining ground in Brazil (Monte *et al.*, 2019). With the intensification of the problems found in chemical resistance breakdown, the use of microorganisms in agriculture is showing promise. It results and is becoming the target of large companies that work with microorganisms for biological control and promoters of plant growth (Ministério da Agricultura, Pecuária e Abastecimento, 2019).

The promotion of plant growth by soil microorganisms can be done through both direct and indirect mechanisms. The direct ones can be the production of hormones or other substances analogous to these, which influence the growth or development of the plant (Zeilinger *et al.*, 2016) or even meeting the required nutritional needs by solubilizing phosphates (Contreras-Cornejo *et al.*, 2016). In contrast, the indirect mechanisms may be through the suppression of pathogens by the action of microorganisms (Saravanakumar *et al.*, 2016; Das *et al.*, 2017; Woo & Pepe, 2018; Mendoza-Mendoza *et al.*, 2018; Monte *et al.*, 2019).

The soil microbiome has the potential to promote plant growth and the tolerance of the plant organism to parasitism of phytopathogens, which may represent a promising sustainable solution to improve agricultural production. Several soil microorganisms, such as fungi, have been reported for their ability to solubilize different forms of inorganic phosphates (Chagas *et al.*, 2016a; Chagas *et al.*, 2017a). Thus, fungi of the *Trichoderma* genus are found naturally in the soil, which has been the most studied and used as an active ingredient in bio fungicides (Karaoglu *et al.*, 2018; Alekseeva *et al.*, 2019), and they also have activity as promoters of plant growth (Chagas *et al.*, 2017b).

Moreover, *Trichoderma* is a rhizosphere fungus that promotes plant growth and can stimulate the plant defense system to suppress attacks by phytopathogens (Rubio *et al.*, 2017; Poole *et al.*, 2018). *Trichoderma* spp.-based bioproducts, such as biofungicides or biofertilizers, are widely used worldwide (Egamberdieva *et al.*, 2017). The influence of *Trichoderma* species on plant development is broad; it includes beneficial effects on seed germination, seedling emergence, grain growth, and productivity (Chagas *et al.*, 2016b; Chagas *et al.*, 2017c; Chagas Junior *et al.*, 2019a; Chagas Junior *et al.*, 2019b).

Therefore, producing low-cost inoculants based on plant growth-promoting microorganisms such as *Trichoderma* spp. is an alternative to reduce the environmental risks caused by the inappropriate and sometimes excessive use of inputs and pesticides. In addition, these fungi increase agricultural production, make the product more competitive and differentiated, and reduce producer costs. Growth-promoting microorganisms can be a great tool for the challenge of modern agriculture in the coming years, which aims to increase sustainable food production with proper environmental protection. Thus, this work aimed to evaluate the efficiency of the product Trichoplus JCO, which contains the active ingredient *Trichoderma asperellum* (Samuels *et al.*, 1999) as a promoter of soybean plant growth.

## MATERIALS AND METHODS

**Experiment location.** Four independent experiments were carried out in different periods between 2019 and 2020. The experiments were conducted in a greenhouse (photoperiod of approximately 12 hours, average temperature of 26°C and humidity of approximately 85%) at the experimental station of the Federal University of Tocantins, Tocantins, Brazil (11°48'29"S, 48°56'39"W, 280m altitude). The geographical coordinates of the experimental station correspond to 11°43'45"S and 49°04'07"W, with an average altitude of 280 meters. The local climatic characterization is of humid tropical climate and classified with minor water deficiency (B1wA'a') with tropical savanna vegetation according to Köppen-Geiger (Peel *et al.*, 2007).

**Product, formulation and *Trichoderma*.** The powdered product Trichoplus JCO (JCO Fertilizers) is a product in a new formulation, containing the active ingredient based on *Trichoderma asperellum* (Concentration of  $2 \times 10^8$  CFU g<sup>-1</sup>), and graphite was used as an inert along with the *Trichoderma* strain cultivated in the laboratory and the product prepared in a production room with controlled temperature and humidity. The molecular identification of the isolate was: *Trichoderma asperellum* GJS 04-217, access to GenBank DQ381958 (99% similarity) (Samuels *et al.*, 2010).

**Experiments.** Experiment 1 was carried out from August to December 2019 in pots with a capacity of 3.8L, filled with soil classified as dystrophic Red-Yellow Latosol (sieved) with a medium texture. The soil was obtained at the UFT Experimental Station at a depth of 0-20cm, with the chemical analysis as shown in Table 1. The granulometric characteristics were 69.7, 7.0 and 23.3% of sand, silt, and clay, respectively (Embrapa - Empresa Brasileira de Pesquisa Agropecuaria, 2011).

**Table 1.** Chemical and organic matter analysis of soil samples used in the experiments.

Experiments	pH	P	K	Al <sup>3+</sup>	H+Al	Ca <sup>2+</sup>	Mg <sup>2+</sup>	SB	T	V	OM
	H <sub>2</sub> O	mg dm <sup>-3</sup>			-----cmol <sub>c</sub> dm <sup>-3</sup> -----					%	%
1	5.1	5.0	9.0	0.0	1.8	0.5	0.2	0.72	2.52	29.0	1.0
2	5.8	5.7	19.0	0.0	1.5	2.2	1.1	3.3	4.9	69	1.5
3	6.5	18.8	30.0	0.0	1.2	7.1	5.6	12.7	13.9	91	10.9
4	5.8	5.7	19.0	0.0	1.5	2.2	1.1	3.3	4.9	69	1.5

Average values of three soil samples collected for each experiment. Chemical attributes of 0-20 cm depth; pH in water - Ratio 1:2.5; P and K - Mehlich 1 extractor; Al<sup>3+</sup>, Ca<sup>2+</sup> and Mg<sup>2+</sup> - KCl extractor (1 mol L<sup>-1</sup>); H + Al - SMP extractor; SB = Sum of exchangeable bases; (T) = Cation Exchange capacity at pH 7.0; V - Base saturation index; and OM = organic matter (oxidation: Na<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub> 4N + H<sub>2</sub>SO<sub>4</sub> 10N).

Experiment 2 was carried out from December 2019 to February 2020 in pots with a capacity of 1.7L, filled with soil classified as dystrophic Red-Yellow Latosol (sieved) with a medium texture. The soil was obtained at the UFT Experimental Station at a depth of 0-20cm in an area other than the soil collected for Experiment 1, which presented the following chemical analysis as shown in Table 1. The granulometric characteristics were 69.0, 7.8 and 23.2% sand, silt, and clay, respectively (Embrapa, 2011).

Experiment 3 was carried out from February to April 2020 in pots with a capacity of 3.8 L, filled with soil classified as haplic plinthosol. The soil was obtained in lowland areas in Formoso do Araguaia, Tocantins, Brazil (11°47'45" S e 49°31'43" W, 240 meters altitude), at a depth of 0-20 cm, with the chemical analysis as shown in Table 1. The granulometric characteristics were 72.5, 10.0 and 17.5% sand, silt, and clay, respectively (Embrapa, 2011).

Experiment 4 was carried out from February to April 2020 in pots with a capacity of 3.8L, filled with soil classified as Red-Yellow Latosol (sieved) with a medium texture. The soil was obtained at the UFT Experimental Station at a depth of 0-20 cm in an area different from the soil collected for experiments 1 and 2, with the chemical analysis shown in Table 1. The granulometric characteristics were 79.8, 8.1 and 12.1% of sand, silt, and clay, respectively (Embrapa, 2011).

In all experiments, essential fertilization (N-P-K + micronutrients) was carried out on the soil before planting, corresponding to 250 kg ha<sup>-1</sup> of formulation 05-25-15.

**Treatments and design of experiments.** Four treatments with different doses of TrichoPlus (3, 4, 5, and 6 g per kg of seeds) and absolute control (without inoculation, negative control) were used in experiment 1. The doses used were considered with the amount of graphite used in soybeans seeds, which has the sole objective of reducing the friction of the seeds with the sowing distribution mechanisms. In contrast, two treatments were used in experiment 2, one with the inoculation of TrichoPlus JCO (5g per kg of seeds) and absolute control (without inoculation).

Inoculation was performed by adding the TrichoPlus JCO product to the seeds in a paper bag and mixing. Then, three treatments were used each in experiments 3 and 4, one with the inoculation of TrichoPlus JCO (5g per kg of seeds), one with the inoculation of commercial product based on *T. asperellum* (positive control), and an absolute control (without inoculation). According to the manufacturer's recommendations, a commercial product at a concentration of  $1 \times 10^{10}$  CFU g<sup>-1</sup> was used for the positive control, a dispersible granular formulation, and a dosage of 100 g per 100 kg of seeds.

The experimental design for the four experiments was entirely randomized, with five treatments and four replicates. In experiments 1, 2 and 3, two evaluations were carried out, and three evaluations were carried out in experiment 4, with four replicates for each evaluation in each experiment. In all experiments, the experimental units were pots with two plants per pot. The soybean seeds were inoculated with the bacterial rhizobia *Bradyrhizobium* (SEMIA 5079 and SEMIA 5080, concentration of  $3 \times 10^9$  CFU mL<sup>-1</sup>) with a dosage of 400mL (four doses of 100 mL, according to the manufacturer) per 50kg seeds in the four experiments before inoculation with TrichoPlus JCO, to supply the nitrogen needed by plants.

**Soybean cultivars and sowing.** The soybean cultivar M 9056 RR was used in experiment 1. The soybean cultivar M 8644 IPRO was used in experiment 2. The soybean cultivar M 8349 IPRO was used in experiments 3 and 4. These cultivars were used in the experiments because they are the most commonly used in the region, being acquired in the region itself. Five seeds per pot were sown in all experiments. The emergence occurred on the fourth day after sowing, and thinning was done, leaving two plants per pot. Daily irrigation was performed according to the field capacity of the pots.

**Evaluations of soybean plants.** In experiment 1, evaluations were made at 25 and 50 days after planting (DAP). For experiment 2, evaluations were made at 15 and 30 DAP. In experiment 3, an evaluation was performed at 40 DAP. For experiment 4, evaluations were performed at 25, 60, and 112 DAP. Height in centimeters, root volume (VR), and biomass were determined using a graduated ruler to measure the height and the root volume in mL by the breaker method. For biomass, the collected

material was washed in running water and taken to dry in an oven at 60°C to determine the dry shoot mass (SDM), root dry mass (RDM), and total dry mass (TDM). The relative efficiency (RE) of each treatment was determined using data from SDM, RDM and TDM according to the equation  $RE = (SDM, RDM, \text{ and } TDM \text{ inoculated with } Trichoderma / SDM, RDM, \text{ and } TDM \text{ without inoculation}) \times 100$ .

The number of nodules (NN), nodules dry mass (NDM), and nitrogen content in the shoot were also determined. SDM at 50 DAP was ground in a knife mill, where a sample was taken to determine the nitrogen content in the shoot by the Kjeldahl method (Malavolta *et al.*, 1997). The nitrogen accumulation in the shoot (NAS) was calculated by multiplying the SDM by the N content. The phosphorus content in the shoot was also determined through nitric-perchloric digestion (Malavolta *et al.*, 1997). The accumulation of P in the tissues of the shoot was determined by multiplying the SDM by its content in the leaf tissue.

**Statistical analysis.** The data were subjected to analysis of variance (ANOVA) and T-test and the averages were compared using the Tukey test at 5% probability, using the SISVAR software version 5.1 statistical analysis program.

## RESULTS AND DISCUSSIONS

**Experiment 1.** The results with the different doses of Trichoplus showed that there were significant differences ( $p < 0.05$ ) being superior for RV and TDM when compared to the control treatment at 25 days after planting (DAP). Among these, 5g kg<sup>-1</sup> was greater (Table 2).

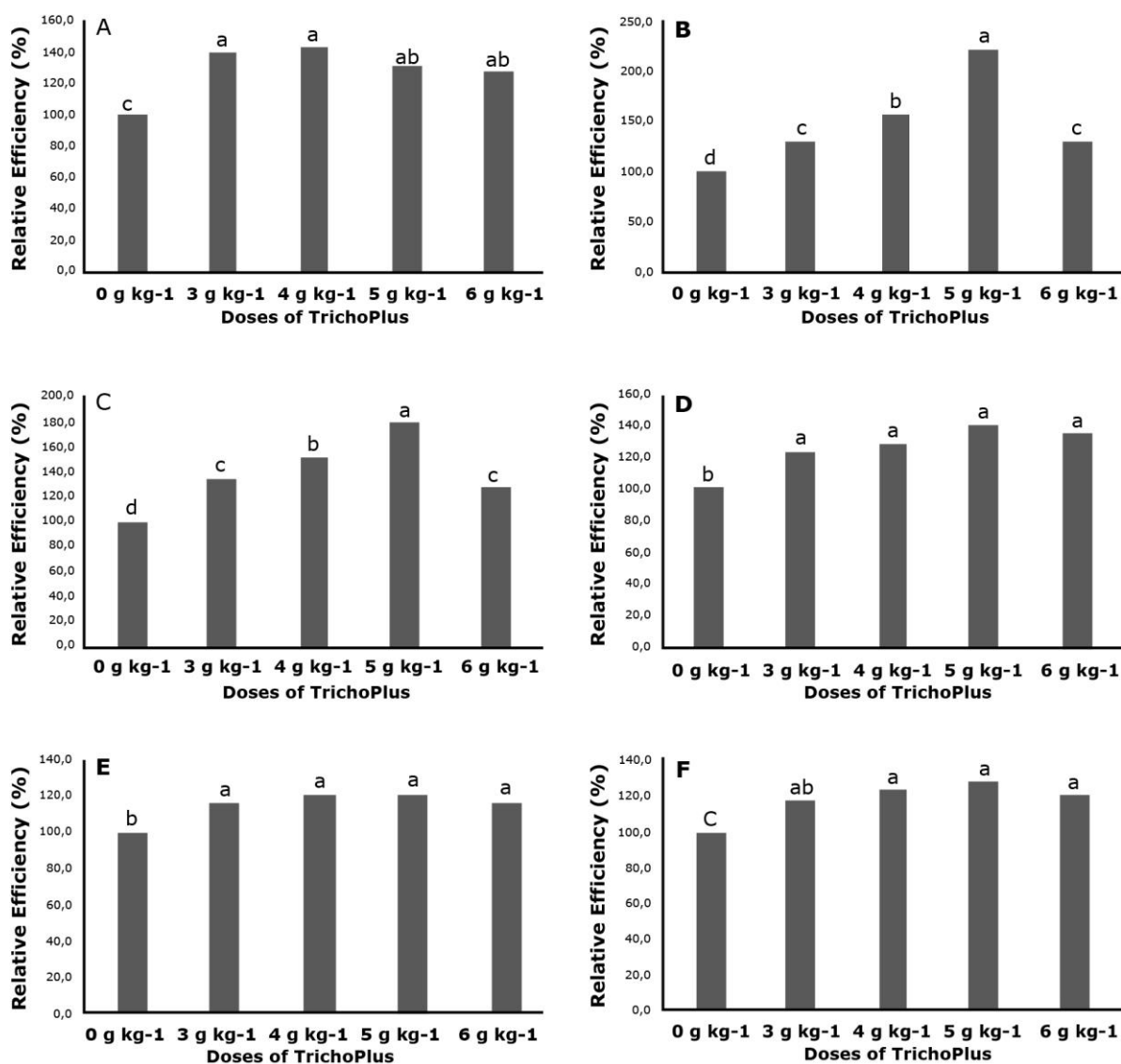
**Table 2.** Biomass of soybean plants cv. M 9056 RR, at 25 and 50 days after planting, inoculated with different doses of TrichoPlus (*Trichoderma asperellum*) applied as in seed treatment.

Treatments	RV cm <sup>3</sup>	SDM g	RDM g	TDM g
25 DAP				
0 g kg <sup>-1</sup>	1.6 c	0.39 b	0.40 b	0.79 c
3 g kg <sup>-1</sup>	2.2 b	0.55 a	0.52 b	1.07 b
4 g kg <sup>-1</sup>	2.7 b	0.56 a	0.63 b	1.19 b
5 g kg <sup>-1</sup>	4.3 a	0.51 a	0.89 a	1.40 a
6 g kg <sup>-1</sup>	2.6 b	0.50 a	0.51 b	1.01 b
CV (%)	16.6	11.3	21.2	11.5
50 DAP				
0 g kg <sup>-1</sup>	5.5 c	0.84 c	1.35 b	2.19 c
3 g kg <sup>-1</sup>	7.5 a	1.05 ab	1.57 a	2.57 ab
4 g kg <sup>-1</sup>	6.9 ab	1.08 ab	1.63 a	2.71 a
5 g kg <sup>-1</sup>	7.4 a	1.17 a	1.64 a	2.81 a
6 g kg <sup>-1</sup>	6.8 ab	1.13 a	1.58 a	2.64 a
CV (%)	6.6	5.3	7.2	8.5

Means followed by the same letter in the column for a DAP do not differ statistically from each other by the Tukey test at the level of 5% probability. CV: Coefficient of Variation. RV, root volume; SDM, shoot dry mass (g); RDM, root dry mass (g); TDM, total dry mass (g).

For the SDM, all treatments with different doses of TrichoPlus were superior to the control. For the RDM, the treatment with the dose of 5g kg<sup>-1</sup> was superior to the others. For the evaluation, at 50 DAP, the RV, SDM, RDM, and TDM, the different doses of TrichoPlus were significantly superior in relation to the control.

For relative efficiency (RE), at 25 DAP, the results were significant ( $p < 0.05$ ) for treatments with different doses of TrichoPlus, with averages ranging from 28.2 to 43.6% for SDM (Figure 1A), between 22.5 to 57.5% for RDM (Figure 1B), and between 27.8 to 77.2 for TDM (Figure 1C) in relation to absolute control without inoculation.



A: shoot dry mass (SDM), B: root dry mass (RDM) and C: total dry mass (TDM) at 25 days after planting; and D: shoot dry mass (SDM), E: root dry mass (RDM) and F: total dry mass (TDM) at 50 days after planting. Means followed by the same letter do not differ by 5% Tukey test.

**Figure 1.** Relative efficiency of soybean plants cv. M 9056 RR inoculated with

TrichoPlus (*Trichoderma asperellum*) concerning to the control without inoculation. Thus, the treatment with 5 g kg<sup>-1</sup> was significantly superior to the other doses at 25 DAP (Figure 1B and Figure 1C). On the other hand, at 50 DAP, superior RE values ( $p < 0.05$ ) were observed for treatments with inoculation of TrichoPlus doses concerning the control, with SDM averages between 25.0 to 39.3% (Figure 1D), RDM averages between 16.3 to 20.7% (Figure 1E), and TDM averages between 17.4 to 28.3% (Figure 1F), in relation to absolute control without inoculation.

The NN was significant ( $p < 0.05$ ) in treatments with 4, 5, and 6 g kg<sup>-1</sup>. For NDM, all treatments with different doses of TrichoPlus were superior ( $p < 0.05$ ) compared to the control (Table 3). In addition, the Nitrogen (N) content and NAS were significantly superior ( $p < 0.05$ ) for the treatments with TrichoPlus in different doses concerning the control. Moreover, the phosphorus content (P) in shoots was superior ( $p < 0.05$ ) in the treatments with the different doses of TrichoPlus and the increased phosphorus content (%P) ranged from 12.9 to 19.4% compared to the control (Table 3). The PAS was significant ( $p < 0.05$ ) in all treatments involving different doses of TrichoPlus. The PAS (4.33g pot<sup>-1</sup>) and % PAS (66.5%) were significantly superior at dose 5g kg<sup>-1</sup> concerning the control.

**Table 3.** Nutrient contents in soybean cultivar M 9056 RR at 50 days after planting inoculated with Trichoplus (*Trichoderma asperellum*).

	NN	NDM	N	NAS	P	%P	PAS	%PAS
0 g kg <sup>-1</sup>	38.5 b	234 a	14.1 b	11.8 c	3.1 b	100 c	2.60 c	100 c
3 g kg <sup>-1</sup>	45.0 ab	267 a	17.8 a	18.7 ab	3.5 a	112.9 ab	3.68 b	141.5 b
4 g kg <sup>-1</sup>	48.0 a	260 a	18.0 a	19.4 a	3.7 a	119.4 a	3.99 ab	153.5 ab
5 g kg <sup>-1</sup>	50.3 a	281 a	18.9 a	22.1 a	3.7 a	119.4 a	4.33 a	166.5 a
6 g kg <sup>-1</sup>	49.5 a	270 a	17.7 a	20.0 a	3.6 a	116.1 a	4.07 ab	156.5 ab
CV (%)	11.6	11.5	10.2	11.0	9.9	9.1	10.8	10.1

Means followed by the same letter do not differ statistically by the Tukey test at 5% probability. CV: Coefficient of Variation. NN, number of nodules; NDM, dry nodule mass (g); N, nitrogen content (g kg.pot<sup>-1</sup>); NAS, nitrogen accumulation in the shoot (g pot<sup>-1</sup>); P, phosphorus content in the shoot (g kg<sup>-1</sup>), %P, phosphorus content increase; PAS, phosphorus accumulation in the shoot (g pot<sup>-1</sup>); %PAS, increase of phosphorus accumulation in the shoot.

**Experiment 2.** The evaluations at 15 and 30 DAP showed that the treatment with TrichoPlus inoculation was superior to control for all biomass parameters (SDM, RDM, and TDM) (Table 4). Likewise, all data obtained from the RE of the biomass analyzes were significant ( $p < 0.05$ ) for the treatment with TrichoPlus inoculation for both 15 and 30 DAP (Figure 2).

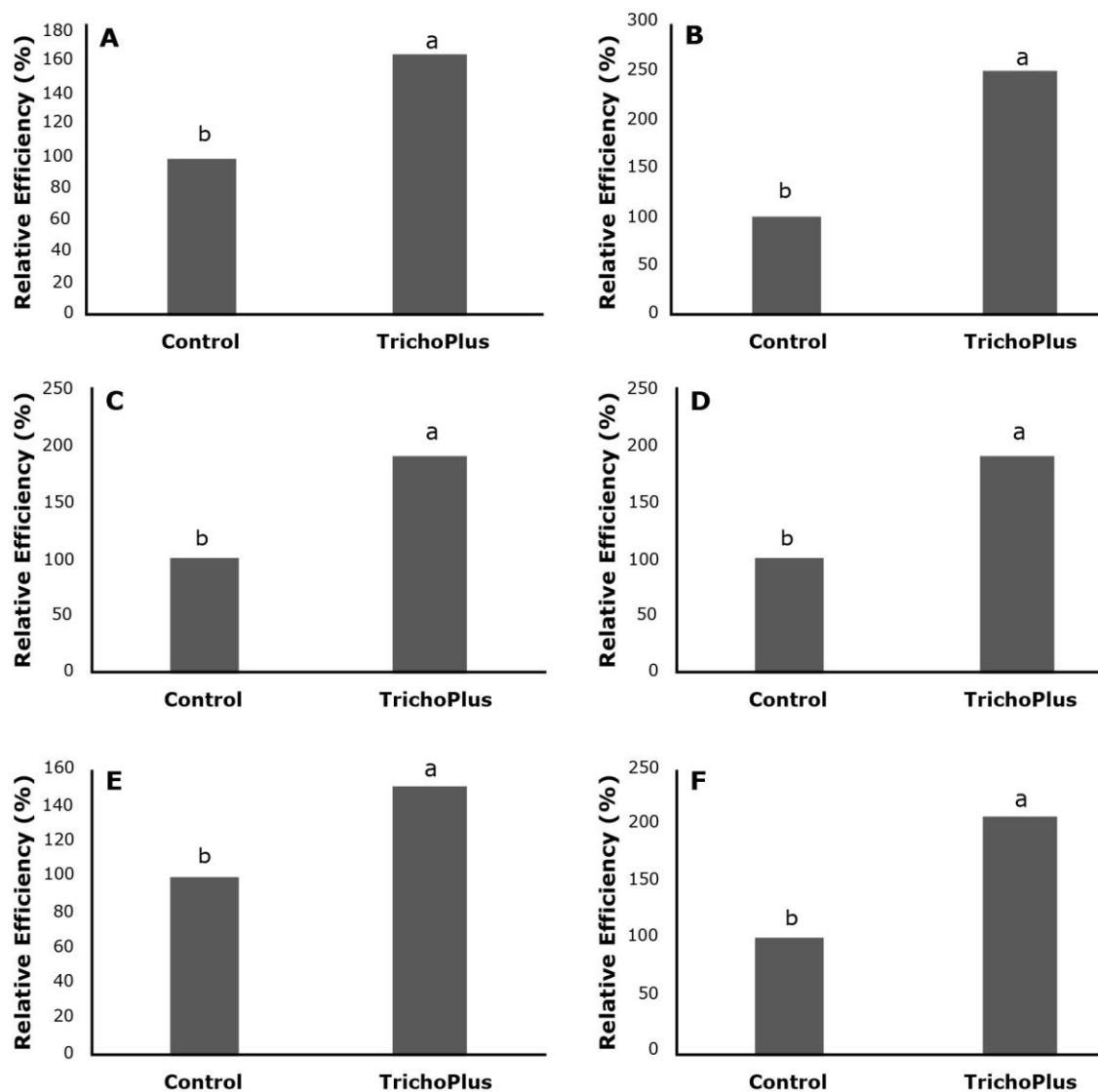
The SDM, RDM, and TDM averages were 64.1, 150, and 91.5% superior to the control, respectively, at 15 DAP. These averages were also 91, 49.6, and 102.2% superior to the control, respectively, at 30 DAP.



**Table 4.** Biomass of soybean plants cv. 8644 IPRO, at 15 and 30 days after planting, inoculated by TrichoPlus (*Trichoderma asperellum*).

Treatments	SDM	RDM	TDM
15 DAP			
TrichoPlus	1.05 a	0.75 a	1.80 a
Control	0.64 b	0.30 b	0.94 b
CV (%)	19.2	23.4	17.8
30 DAP			
TrichoPlus	2.77 a	1.69 a	3.66 a
Control	1.45 b	1.13 b	1.81 b
CV (%)	18.3	17.9	13.9

Means followed by the same letter in the column for a DAP do not differ statistically from each other by the Tukey test at 5% probability. CV: Coefficient of Variation. SDM, shoot dry mass (g); RDM, root dry mass (g); TDM, total dry mass (g).



A: shoot dry mass (SDM), B: root dry mass (RDM) and C: total dry mass (TDM) at 15 days after planting; and D: shoot dry mass (SDM), E: root dry mass (RDM) and F: total dry mass (TDM) at 30 days after planting. Means followed by the same letter do not differ by 5% Tukey test.

**Figure 2.** Relative efficiency of soybean plants cv. 8644 IPRO inoculated with TrichoPlus (*Trichoderma asperellum*) compared to the control without inoculation.

**Experiment 3.** The treatment with Trichoplus was significantly superior ( $p < 0.05$ ) for SDM, RDM, and TDM concerning the positive control (commercial product) and the absolute control (without inoculation) (Table 5). Otherwise, NN and NDM had no significant differences between treatments.

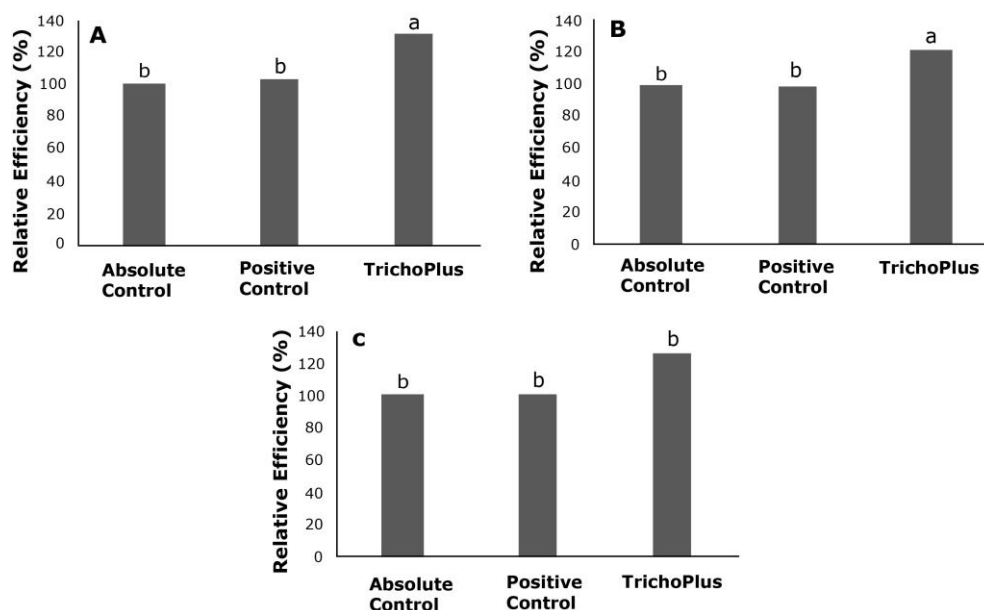
**Table 5.** Biomass and nutrient contents in soybean plants cv. 8349 IPRO inoculated with TrichoPlus (*Trichoderma asperellum*) at 40 days after planting.

Treatments	SDM	RDM	TDM	NN	NDM	N	NAS
TrichoPlus	3.95 a	3.55 a	7.50 a	28.5 a	214 a	15.9 a	62.8 a
Positive control	3.10 b	2.90 b	6.00 b	25.3 a	210 a	14.7 a	45.6 b
Absolute control	3.00 b	2.95 b	5.95 b	26.5 a	201 a	11.4 b	34.2 c
CV (%)	7.5	14.5	9.1	11.6	11.5	13.2	13.1

Means followed by the same letter in the column do not differ statistically from each other by the Tukey test at 5% probability. CV: Coefficient of Variation. Shoot dry mass (SDM, g), root dry mass (RDM, g), total dry mass (TDM, g), number of nodules (NN), dry nodule mass (NDM, g), nitrogen content (N, g kg.pot<sup>-1</sup>) and nitrogen accumulation in the shoot (NAS, g pot<sup>-1</sup>).

The treatments with TrichoPlus and the positive control were significantly superior in N content to the absolute control. In addition, the NAS was significantly superior ( $p < 0.05$ ) for the treatment with TrichoPlus inoculation concerning the positive control and absolute control treatments.

The relative efficiencies were significant ( $p < 0.05$ ) in the SDM (28.4 and 31.7%), RDM (22 and 20.3%), and TDM (25.3 and 26.1%) for the treatment inoculated with TrichoPlus compared to the positive and absolute control (Figure 3).



A: shoot dry mass (SDM), B: root dry mass (RDM), and C: total dry mass (TDM). Means followed by the same letter do not differ by 5% Tukey test.

**Figure 3.** Relative efficiency of soybean plants cv. M 8349 IPRO inoculated with TrichoPlus (*Trichoderma asperellum*) compared to the positive control (commercial product) and absolute control (without inoculation) at 40 days after planting.

On the other hand, the P content in the shoot of the treatment with TrichoPlus and the positive control was superior ( $p < 0.05$ ) with an increase in %P of 45 and 30%, respectively, compared to the absolute control (Table 6). Then, the PAS and %PAS were superior with treatment inoculated by TrichoPlus, with 92% relative to the absolute control and 57% to the positive control.

**Table 6.** Nutrient contents in the shoot of soybean plants cv. M 8349 IPRO inoculated with TrichoPlus (*Trichoderma asperellum*) compared to a positive control (commercial inoculant) and absolute control (without inoculation).

Treatments	P	%P	PAS	%PAS
TrichoPlus	2.9 a	145 a	11.5 a	192 a
Positive control	2.6 a	130 a	8.1 b	135 b
Absolute control	2.0 b	100 b	6.0 bc	100 c
CV (%)	9.4	9.1	10.8	10.5

Means followed by the same letter in the column do not differ statistically from each other by the Tukey test at 5% probability. CV: Coefficient of variation. P, phosphorus content ( $\text{g kg}^{-1}$ ); %P, phosphorus content increase; PAS, phosphorus accumulation in the shoot ( $\text{g pot}^{-1}$ ); %PAS, increase in phosphorus accumulation in the shoot.

**Experiment 4.** In this experiment, the height in the treatment with TrichoPlus and in the positive control was significantly ( $p < 0.05$ ) than the absolute control at 25 DAP (Table 7). The RV, RDM, and TDM in the treatment with TrichoPlus were significantly superior ( $p < 0.05$ ) to the positive control (commercial product) and the absolute control (without inoculation). Otherwise, the SDM in the treatment with TrichoPlus only differed significantly against the absolute control.

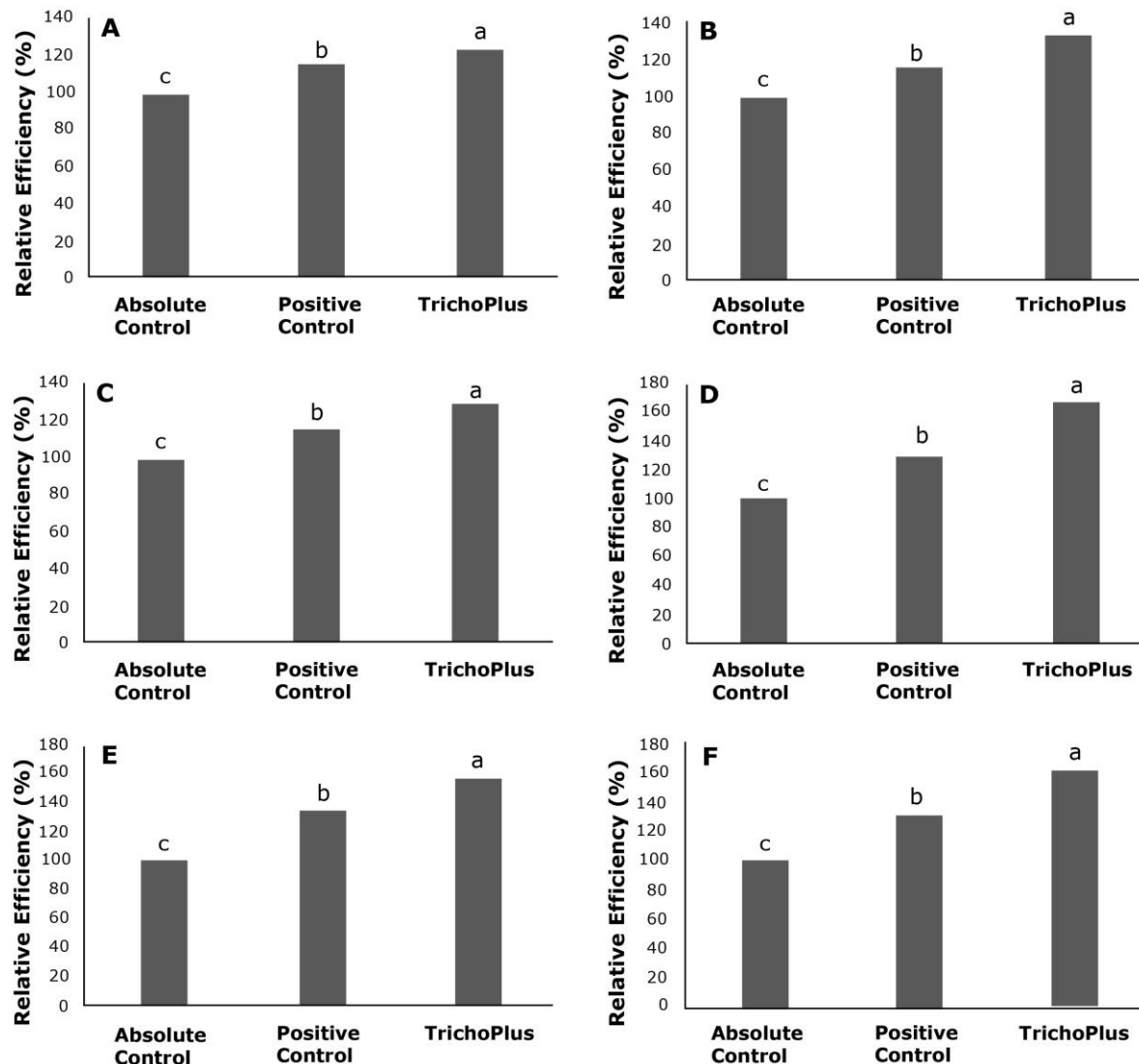
Moreover, the plant heights did not show significant differences between treatments at 60 DAP. The RV in the treatment with TrichoPlus and the positive control was superior ( $p < 0.05$ ) than that in the absolute control. The SDM, RDM, and TDM were superior in the treatment with TrichoPlus concerning the positive control (commercial product) and the absolute control (without inoculation).

**Table 7.** Biomass of soybean plants cv. M 8349 IPRO inoculated with Trichoplus (*Trichoderma asperellum*) at 25 and 60 days after planting (DAP).

Treatments	Height	RV	SDM	RDM	TDM
25 DAP					
TrichoPlus	11.53 a	7.08 a	0.99 a	0.80 a	1.79 a
Positive control	10.78 a	6.15 b	0.93 ab	0.69 b	1.62 b
Absolute control	8.45 b	5.53 c	0.81 b	0.60 b	1.41 c
CV (%)	7.29	4.73	6.99	6.24	4.01
60 DAP					
TrichoPlus	34.0 a	12.1 a	5.23 a	3.73 a	8.96 a
Positive control	33.5 a	12.0 a	4.05 b	3.20 b	7.25 b
Absolute control	33.0 a	8.3 b	3.13 c	2.38 c	5.51 c
CV (%)	4.12	10.22	10.87	7.86	6.48

Means followed by the same letter in the column for a DAP do not differ statistically from each other by the Tukey test at 5% probability. CV: Coefficient of variation. Height (cm); RV, root volume, SDM, shoot dry mass (g); RDM, root dry mass (g); TDM, total dry mass (g).

At 25 DAP, the relative efficiency values for the treatment with TrichoPlus in SDM was 22.2% and 7.4% higher ( $p < 0.05$ ) than the absolute and positive controls (Figure 4). Likewise, RDM relative efficiency values with TrichoPlus were 33.3 and 18.3% superior ( $p < 0.05$ ) than the absolute control and positive control treatments, respectively. TDM relative efficiency values with TrichoPlus were 27.0 and 12.1% superior ( $p < 0.05$ ) than the absolute and positive controls treatments, respectively. Moreover, SDM (67.1 and 37.7%), RDM (56.7 and 22.2%), and TDM (62.6 and 31.0%) relative efficiency values with TrichoPlus treatment were superior to the absolute control and positive control treatments, respectively, at 60 DAP.



A: shoot dry mass (SDM), B: root dry mass (RDM), and C: total dry mass (TDM) at 25 DAP; D: shoot dry mass (SDM), E: root dry mass (RDM), and F: total dry mass (TDM) at 60 DAP. Means followed by the same letter do not differ by 5% Tukey test.

**Figure 4.** Relative efficiency of soybean plants cv. M 8349 IPRO inoculated with TrichoPlus (*Trichoderma asperellum*) compared to the positive control (commercial product) and absolute control (without inoculation).

The analysis of the nodules showed that the NN and NDM with TrichoPlus treatment were significantly superior ( $p < 0.05$ ) to the positive and absolute controls (Table 8). The N content had no significant differences between treatments. However, the NAS with TrichoPlus treatment was significant ( $p < 0.05$ ) concerning the positive control and absolute control treatments. On the other hand, the phosphorus (P) content with TrichoPlus treatment and the positive control were superior ( $p < 0.05$ ) with a P content increase (%P) of 36.7 and 26.7%, respectively, compared to absolute control. Thereafter, PAS and %PAS with TrichoPlus treatment were superior ( $p < 0.05$ ) than both controls, which showed an increase in %PAS with TrichoPlus of 128% concerning the absolute control and 64% in relation to positive control.

**Table 8.** Biomass and nutrient contents in soybean cv. M 8349 IPRO 60 days after planting inoculated with TrichoPlus (*Trichoderma asperellum*).

Treatments	NN	NDM	N	NAS	P	%P	PAS	%PAS
TrichoPlus	42.5 a	314 a	17.4 a	91.0 a	4.1 a	137 a	21.4 a	228 a
Positive control	35.3 b	260 b	16.2 ab	65.6 b	3.8 a	127 ab	15.4 b	164 b
Absolute control	36.0 b	271 b	15.8 ab	49.5 c	3.0 b	100 c	9.4 c	100 c
CV (%)	11.7	11.5	10.5	10.2	6.6	6.3	7.5	7.2

Means followed by the same letter in the column do not differ statistically from each other by the Tukey test at 5% probability. CV: Coefficient of variation. NN, number of nodules; NDM, nodules dry mass (g); N, nitrogen content ( $\text{g kg}^{-1}$ ); NAS, nitrogen accumulation in shoots ( $\text{g pot}^{-1}$ ); P, phosphorus content ( $\text{g kg}^{-1}$ ); %P, phosphorus content increase; PAS, phosphorus accumulation in shoots ( $\text{g pot}^{-1}$ ); %PAS, increase in phosphorus accumulation in shoots.

In the third evaluation, height and grain weight per pot (GWP) obtained with the treatment of TrichoPlus were significant ( $p < 0.05$ ) in relation to both controls (Table 9). The number of pods (NP) and the number of grains (NG) of the treatments with TrichoPlus and positive control were significantly higher ( $p < 0.05$ ) than the absolute control.

**Table 9.** Height and yield of soybean cv. 8349 IPRO inoculated with TrichoPlus (*Trichoderma asperellum*) at 112 days after planting.

Treatments	Height	NI	NP	NG	GWP
TrichoPlus	71.75 a	19.0 a	39.9 a	122.5 a	14.2 a
Positive control	66.13 b	18.9 a	37.8 a	112.5 a	11.3 b
Absolute control	60.88 b	16.6 a	27.0 b	82.3 b	9.2 b
CV (%)	4.59	8.18	7.55	6.25	10.71

Means followed by the same letter in the column do not differ statistically from each other by the Tukey test at 5% probability. CV: Coefficient of variation. Height (cm); NI, number of internodes ( $\text{plant}^{-1}$ ); NP, number of pods ( $\text{plant}^{-1}$ ); NG, number of grains ( $\text{plant}^{-1}$ ); GWP, grain weight per pot ( $\text{g pot}^{-1}$ ).

All of these effects may be related to the ability of *Trichoderma* to promote plant growth, as already reported in the literature (Contreras-Cornejo *et al.*, 2015; Li *et al.*, 2015; Contreras-Cornejo *et al.*, 2016; Monte *et al.*, 2019), and the production of auxins and metabolites such as 6PP, which favors the development of the roots, becoming more profound and more vigorous (Zeilinger *et al.*, 2016; Rubio *et al.*, 2017; Mendoza-Mendoza *et al.*, 2018). Several fungi species have been reported to produce auxins in

interactions with plants, which are essential hormones that affect growth and development (Contreras-Cornejo *et al.*, 2016; Chagas *et al.*, 2017a). They can also increase the absorption and solubilization of nutrients, such as phosphates, through acidification, redox, chelation, and hydrolysis processes (Li *et al.*, 2015). Likewise, these microorganisms can favor hydrophobic adhesion and the development of absorbent root hairs on the side roots, which increases the water absorption surface and that of some important nutrients, such as nitrogen (Machado *et al.*, 2012; Das *et al.*, 2017; Woo & Pepe, 2018; Mendoza-Mendoza *et al.*, 2018).

The active component of the product TrichoPlus, *T. asperellum*, showed the production of indole acetic acid, and they also promoted the increase in the production of soybean plant biomass in previous studies, which indicates the relationship between the production of hormones and biomass (Chagas *et al.*, 2017a; Chagas Junior *et al.*, 2019a). Thus, the biomass accumulation in soybean plants in this study can be related to the production of hormones or growth factors, greater efficiency in using nutrients such as P, and increased availability and absorption of this nutrient by plants.

In addition, the fungus *T. asperellum* can acidify the environment where it is established by the secretion of organic acids such as gluconic, citric, or fumaric acids simultaneously with the synthesis or stimulation of phytohormone production. These acids are the result of carbon source metabolism, mainly glucose, and are capable of solubilizing phosphates, micronutrients, and cations, including iron, manganese, and magnesium, according to Harman *et al.* (2004). Thus, the addition of *Trichoderma* to soils with a scarcity of these cations could result in their solubilization of the available elements, or the addition of poorly soluble natural phosphates as an alternative to phosphorus supplies in the soil, which would increase the production of biomass and productivity of crops (Contreras-Cornejo *et al.*, 2016). Studies have shown that species of *Trichoderma* spp. can promote increases of up to 300% in plant growth (Brotman *et al.*, 2010). The results obtained in the greenhouse experiments corroborate those obtained by Silva *et al.* (2011), who evaluated the effect of *Trichoderma* on the growth of cucumbers and observed a significant increase concerning the control without inoculation of *Trichoderma*.

In all the experiments, there was an increase in the biomass of the root system, and this increase in the root part is related to the health of the plant provided by the microorganisms. Thus, plants that contain this microorganism associated with their roots or in the rhizosphere tend to have a better ability to survive and absorb nutrients in adverse situations and consequently have a productive advantage over those that do not have *Trichoderma* in their roots (Contreras-Cornejo *et al.*, 2015; Contreras-Cornejo *et al.*, 2016).

This study's results agree with other studies using different agricultural cultures inoculated with different isolates of *Trichoderma*. Santos *et al.* (2010) concluded that the use of *Trichoderma* spp. provided positive results in increasing the fresh and dry mass of passion fruit plants from cuttings. Carvalho *et al.* (2011) evaluated the inoculation of *Trichoderma* isolates in promoting the initial growth of common bean. Jesus *et al.* (2011) highlighted the potential of *T. asperellum* as a substrate conditioner to produce coffee seedlings, proving the positive effect on the increase in the root, shoot, and total biomass, as well as the increase in the efficiency of P absorption. Silva *et al.* (2012) demonstrated that *Trichoderma* isolates obtained from Amazonian soils increased the biomass of rice plants in the greenhouse, showing potential as growth promoters. Machado *et al.* (2012) also pointed out that research shows that *Trichoderma* spp. is efficient, practical, and safe in terms of application methods, biocontrol and plant growth promotion. Likewise, studies have shown the positive effect of *Trichoderma* on cowpea and soybeans grown in a greenhouse and field conditions (Chagas *et al.*, 2016a; Chagas *et al.*, 2016b; Chagas Junior *et al.*, 2019a; Chagas Junior *et al.*, 2019b).

The results obtained with the application of the inoculant developed with the *Trichoderma asperellum* strain, which makes up the TrichoPlus product, are promising, especially for the culture under study. The success of the preparation of the inoculant and the inoculation can be credited to its effect as a promoter of plant growth. The benefits of ecological processes performed by fungi of the *Trichoderma* genus as promoters of plant growth have contributed to achieving sustainability in the agricultural sector (Meyer *et al.*, 2020; Meyer *et al.*, 2022).

The inoculation technology of microorganisms such as the fungus *Trichoderma* of biotechnological interest in agriculture is a resource of great economic importance, in addition to the contribution to the reduction of use and the consequent impact of agrochemicals, and the global market of biologicals for agriculture, which involves biopesticides, inoculants, biostimulants, and biofertilizers, was estimated at US\$ 9.9 billion in 2020 (Borsari & Vieira, 2022).

Sustainable agriculture requires the use of a strategy that increases productivity without harming the environment, opening up new perspectives to contribute to the development of new technologies, methods, and strategies in agribusiness. The processes mediated by microorganisms become essential in preserving and conserving natural resources.

In addition, biological products based on *Trichoderma* are growing as an alternative to the use of chemical products, being considered to have a low impact on the environment and providing healthier food to the population (Chagas Junior *et al.*,

2018). These products also appear as an important alternative in the biocontrol of phytopathogens due to the different mechanisms of action such as competition (water, air, light, and nutrients); host-parasite association, which can be physical or metabolic with digestion by hydrolytic enzymes, including chitinases, proteases, glucanases, and lipases; and antibiosis, which is the action of a microorganism that acts on another, releasing substances that harm or kill another microorganism (Woo *et al.*, 2014; Oliveira *et al.*, 2022).

The information reported in this study, with the inoculation of TrichoPlus in this formulation, inoculated in soybean seeds, and in a greenhouse, need to be tested in applications under field conditions to prove the efficiency of *Trichoderma* as a plant growth promoter.

## CONCLUSION

The inoculation of TrichoPlus, a *Trichoderma asperellum*-containing product, in the dosage of 5 g per kg of seed resulted in significantly better values for the biomass accumulation, contents of nitrogen and phosphorus in soybeans. Such findings demonstrate that TrichoPlus can be recommended as an efficient inoculant, promoting the plant growth of soybean plants in the greenhouse.

## ACKNOWLEDGMENTS

We are grateful to the Graduate Program in Plant Production at the Federal University of Tocantins (UFT), Gurupi, TO, Brazil. This work was supported by the partnership between JCO Bioprodutos, UFT and the Foundation for Scientific and Technological Support of Tocantins (FAPTO).

**Conflict of interest:** The authors declare that there is no conflict of interest.

## BIBLIOGRAPHIC REFERENCES

- Alekseeva, K.L.; Smetanina, L.G.; Kornev, A.V. (2019). Biological protection of tomato from *Fusarium* wilt. *AIP Conference Proceedings*. 2063(1): 030001. <https://doi.org/10.1063/1.5087309>
- Borsari, A.C.P.; Vieira, L.C. (2022). Mercado e perspectivas dos bioinsumos no Brasil. In: Meyer, M.C.; Bueno, A.F.; Mazaro, S.M.; Silva, J.C. (Eds.). *Bioinsumos na cultura da soja*. p. 39-52. Brasília: Embrapa.
- Brotman, Y.; Gupta K.J.; Viterbo, A. (2010). *Trichoderma*. *Current Biology*. 20(9): 390-391. [10.1016/j.cub.2010.02.042](https://doi.org/10.1016/j.cub.2010.02.042)
- Carvalho, D.D.C.; Mello, S.C.M.; Lobo Junior, M.; Silva, M.C. (2011). Control of *Fusarium oxysporum* f. sp. *phaseoli* in vitro and on seeds and growth promotion of common bean in early stages by *Trichoderma harzianum*. *Tropical Plant Pathology*. 36(1): 28-34. [10.1590/S1982-56762011000100004](https://doi.org/10.1590/S1982-56762011000100004)



- Chagas, L.F.B.; Castro, H.G.; Colonia, B.S.O.; Carvalho Filho, M.R.; Miller, L.O.; Chagas Junior, A.F. (2016a). Efficiency of *Trichoderma* spp. as a growth promoter of cowpea (*Vigna unguiculata*) and analysis of phosphate solubilization and indole acetic acid synthesis. *Brazilian Journal of Botany*. 38(4): 1-11. 10.1007/s40415-015-0247-6
- Chagas, L.F.B.; Castro, H.G.; Colonia, B.S.O.; Carvalho Filho, M.R.; Miller, L.O.; Chagas Junior, A.F. (2016b). Efficiency of the inoculation of *Trichoderma asperellum* UFT-201 in cowpea production components under growth conditions in field. *Revista de Ciências Agrárias*. 39(3): 413-421. 10.19084/RCA15112
- Chagas, L.F.B.; Chagas Junior, A.F.; Castro, H.G. (2017a). Phosphate solubilization capacity and indole acetic acid production by *Trichoderma* strains for biomass increase on basil and mint plants. *Brazilian Journal of Agriculture*. 92(2): 176-185. 10.37856/bja.v92i2.3221
- Chagas, L.F.B.; Chagas Junior, A.F.C.; Soares, L.P.; Fidelis, R.R. (2017b). *Trichoderma* na promoção do crescimento vegetal. *Revista de Agricultura Neotropical*. 4(3): 97-102. <https://doi.org/10.32404/rean.v4i3.1529>
- Chagas, L.F.B.; Martins, A.L.L.; Carvalho Filho, M.R.; Miller, L.O.; Oliveira, J.C.; Chagas Junior, A.F. (2017c). *Bacillus subtilis* e *Trichoderma* spp. no incremento da biomassa em plantas de soja, feijão-caupi, milho e arroz. *Agri-Environmental Sciences*. 3(2): 10-18. <https://revista.unitins.br/index.php/agri-environmental-sciences/article/view/430>
- Chagas Junior, A.F.C.; Chagas, L.F.B.; Santos, G.R.; Martins, A.L.L.; Carvalho F.M.R.; Oliveira-Miller, L. (2018). Action of *Trichoderma* spp. in the control of *Fusarium* sp., *Rhizoctonia solani* and *Sclerotium rolfsii*. *Agri-Environmental Sciences*. 4(2): 9-15. 10.36725/agries.v4i2.420
- Chagas Junior, A.F.; Chagas, L.F.B.; Miller, L.O.; Oliveira, J.C. (2019a). Efficiency of *Trichoderma asperellum* UFT 201 as plant growth promoter in soybean. *African Journal of Agricultural Research*. 14(5): 263-271. 10.5897/AJAR2018.13556
- Chagas Junior, A.F.; Chagas, L.F.B.; Colonia, B.S.O.; Miller, L.O.; Oliveira, J.C. (2019b). *Trichoderma asperellum* (UFT201) functions as a growth promoter for soybean plant. *African Journal of Agricultural Research*. 14(33):1772-1777. 10.5897/AJAR2019.13985
- Contreras-Cornejo, H.A.; López-Bucio, J.S.; Méndez-Bravo, A.; Macías-Rodríguez, L.; Ramos-Vega, M.; Guevara-García, A.A.; López-Bucio, J. (2015). Mitogen-activated protein kinase 6 and ethylene and auxin signaling pathways are involved in *Arabidopsis* root-system architecture alterations by *Trichoderma atroviride*. *Molecular Plant Microbe Interactions*. 28(6): 701-10. <https://doi.org/10.1094/MPMI-01-15-0005-R>
- Contreras-Cornejo, H.A.; Macías-Rodríguez, L.; Del-Val, E.; Larsen, J. (2016). Ecological functions of *Trichoderma* spp. and their secondary metabolites in the rhizosphere: interactions with plants. *FEMS Microbiology Ecology*. 92: 1-17. 10.1093/femsec/fiw036
- Das, T.; Mahapatra, S.; Das, S. (2017). In vitro compatibility study between the *Rhizobium* and native *Trichoderma* isolates from lentil rhizospheric soil. *International Journal of Current Microbiology and Applied Sciences*. 6(8):1757-1769. 10.20546/ijcmas.2017.608.208
- Egamberdieva, D.; Wirth, S.J.; Shurigin, V.V.; Hashen, A.; Allah, E.F.A. (2017). Endophytic bacteria improve plant growth, symbiotic performance of chickpea (*Cicer arietinum* L.) and induce suppression of root rot caused by *Fusarium solani* under salt stress. *Frontiers in Microbiology*. 8:1-13. <https://doi.org/10.3389/fmicb.2017.01887>
- Embrapa-Empresa Brasileira de Pesquisa Agropecuária. (2011). *Manual de métodos de análise de solo*. 2ª edition. Rio de Janeiro: EMBRAPA-CNPQ: 225p.

- Harman, G.E.; Howell, C.R.; Viterbo, A.; Chet, I.; Lorito, M. (2004). *Trichoderma* species-opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*. 2(1): 43-56. <https://doi.org/10.1038/nrmicro797>
- Jesus, E.P.; Souza, C.H.E.; Pomella, A.W.V.; Costa, R.L.; Seixas, L.; Silva, R.B. (2011). Avaliação do potencial de *Trichoderma asperellum* como condicionador de substrato para a produção de mudas de café. *Cerrado Agrociências*. 2: 7-19.
- Karaoglu, S.; Bozdeveci, A.; Pehlivan, N. (2018). Characterization of local *Trichoderma* sp. as potential bio-control agents, screening of *in vitro* antagonistic activities and fungicide tolerance. *Hacettepe Journal of Biology and Chemistry*. 46(2): 247-261. 10.15671/HJBC.2018.233
- Li, R.X.; Cai, F.; Pang, G.; Shen, Q.R.; Li, R.; Chen, W. (2015). Solubilisation of phosphate and micronutrients by *Trichoderma harzianum* and its relationship with the promotion of tomato plant growth. *PLoS One*. 10(6): e0130081. 10.1371/journal.pone.0130081
- Machado, D.F.M.; Parzianello, R.F.; Silva, A.C.F.; Antonioli, Z.I. (2012). *Trichoderma* no Brasil: o Fungo e Bioagente. *Revista de Ciências Agrárias*. 35(1):274-288. 10.19084/rca.16182
- Malavolta, E.; Vitti, G.C.; Oliveira, S.A. (1997). Princípios, métodos e técnicas de avaliação do estado nutricional. In: Malavolta, E.; Vitti, G.; Oliveira, A.S. *Avaliação do estado nutricional de plantas: princípios e aplicações*. 115-230p. 2ª edition. Piracicaba: Potafos.
- Mendoza-Mendoza, A.; Zaid, R.; Lawry, R.; Hermosa, R.; Monte, E.; Horwitz, B.A.; Mukherjee, P.K. (2018). Molecular dialogues between *Trichoderma* and roots: role of the fungal secretome. *Fungal Biology Reviews*. 32(2): 62-85. 10.1016/j.fbr.2017.12.001
- Meyer, M.C.; Mazaro, S.M.; Silva, J.C. (2020). *Trichoderma*: uso na agricultura. Brasília: Embrapa. 538p.
- Meyer, M.C.; Mazaro, S.M.; Silva, J.C. (2022). Bioinsumos na cultura da soja. Brasília: Embrapa. 550p.
- Ministério da Agricultura, Pecuária e Abastecimento. (2019). Mercado de biodefensivos cresce mais de 70% no Brasil em um ano. [www.agricultura.gov.br/noticias/feffmercado-de-biodefensivos-cresce-em-mais-de-50-no-brasil](http://www.agricultura.gov.br/noticias/feffmercado-de-biodefensivos-cresce-em-mais-de-50-no-brasil)
- Monte, B.H.; Bettiol, E.; Hermosa, R. (2019). *Trichoderma* e seus mecanismos de ação para o controle de doenças de plantas. In: Meyer, M.C.; Mazaro, S.M.; Silva, J.C. *Trichoderma: Uso na Agricultura*. 81-199. Brasília: Embrapa.
- Oliveira, R.S.; Chagas, L.F.B.; Martins, A.L.L.; Souza, M.C.; Luz, L.L.; Gomes, F.L.; Chagas Junior, A.F. (2022). *Trichoderma* in the phytopathogenic biocontrol. *Bulgarian Journal of Agricultural Science*. 28(4): 717-724.
- Peel, M.; Finlayson, B.L.; McMahon, T.A. (2007). Update world map of the Köppen-Geiger climate classification. *Hydrology and Earth System Science*. 11: 1633-1644. <https://doi.org/10.5194/hess-11-1633-2007>
- Poole, P.P.; Ramachandran, V.; Terpolilli, J. (2018). Rhizobia: from saprophytes to endosymbionts. *Nature Reviews Microbiology*. 18(5):291-303. 10.1038/nrmicro.2017.171
- Rubio, M.B.; Hermosa, R.; Vicente, R.; Gómez-Acosta, F.A.; Morcuende, R.; Monte, E.; Bettiol, W. (2017). The combination of *Trichoderma harzianum* and chemical fertilization leads to the deregulation of phytohormone networking, preventing the adaptive responses of tomato plants to salt stress. *Frontiers in Plant Science*. 8(294): 1-14. 10.3389/fpls.2017.00294
- Samuels, G.J.; Lieckfeldt, E.; Nirenberg, H.I. (1999). *Trichoderma asperellum*, a new species with warted conidia, and redescription of *T. viride*. *Sydowia*. 51(1): 71-88.

Samuels, G.J.; Ismaiel, A.; Bon, M.C.; De Respinis, S.; Petrini, O. (2010). *Trichoderma asperellum* sensu lato consists of two cryptic species. *Mycologia*. 102(4): 944-966. 10.3852/09-243

Santos, H.A.; Mello, S.C.M.; Peixoto, J.R. (2010). Associação de isolados de *Trichoderma* spp. e ácido indol-3-butírico (AIB) na promoção de enraizamento de estacas e crescimento de maracujazeiro. *Bioscience Journal*. 26(6): 966-972.

Saravanakumar, K.; Yu, C.; Dou, K.; Wang, M.; Li, Y.; Chen, J. (2016). Synergistic effect of *Trichoderma*-derived antifungal metabolites and cell wall degrading enzymes on enhanced biocontrol of *Fusarium oxysporum* f. sp. *Cucumerinum*. *Biological Control*. 94: 37-46. 10.1016/j.biocontrol.2015.12.001

Silva, V.N.; Guzzo, S.D.; Lucon, C.M.M.; Harakava, R. (2011). Promoção de crescimento e indução de resistência à antracnose por *Trichoderma* spp. em pepineiro. *Pesquisa Agropecuária Brasileira*. 46(12): 1609-1618.

Silva, J.C.; Torres, D.B.; Lustosa, D.C.; Filippi, M.C.C.; Silva, G.B. (2012). Rice sheath blight biocontrol and growth promotion by *Trichoderma* isolates from the Amazon. *Amazonian Journal of Agricultural and Environmental Sciences*. 55(4): 243-250. <http://dx.doi.org/10.4322/rca.2012.078>

Woo, S.L.; Pepe, O. (2018). Microbial consortia: promising probiotics as plant biostimulants for sustainable agriculture. *Frontiers*. 9(1801): 1-6.

Woo, S.L.; Ruocco, M.; Vinale, F.; Nigro, M.; Marra, R.; Lombardi, N.; Pascale, A.; Lanzuise, S.; Manganiello, G.; Lorito, M. (2014). *Trichoderma*-based products and their widespread Use in agriculture. *The Open Mycology Journal*. 8: 71-126.

Zeilinger, S.; Gruber, S.; Bansal, R. (2016). Secondary metabolism in *Trichoderma*-Chemistry meets genomics. *Fungal Biology Reviews*. 30(2): 74-90. <https://doi.org/10.1016/j.fbr.2016.05.001>